

Remarks

Claims 1-20 were pending. Claims 10, 11, 13 and 15 are cancelled without prejudice to prosecution in a future application, due to the restriction requirement. Claims 1 and 19 were cancelled as redundant. Claims 21-42 were added. Therefore, claims 2-9, 12, 16-18 and 20-42 are now pending

Amendments to the Claims

Claims 2-9, 12, and 16 were amended. Support for the new and amended claims can be found throughout the specification, for example:

Claims 2, 6, 7, 12: page 2, lines 6-7 and 19-21.
Claim 3: original claim 3.
Claims 4, 5, 9: amended for readability only.
Claim 8: original claim 8 and page 13, lines 22-25.
Claim 16: original claim 8.
Claims 21-32: original claim 3.
Claim 33: original claim 8.
Claim 34: page 17, lines 15-16.
Claims 35-36: page 4, lines 23-28.
Claim 37-38: original claim 3.
Claim 39: page 29, line 25- page 30, line 27.
Claims 40-42: page 32, line 20- page 34, line 21.

No amendments herein were made to distinguish prior art. Instead, the amendments were made to clarify the claims.

Summary of Telephone Interview

Applicants thank Examiner Kruse for the courtesy of a telephone interview with Applicants' representative Sheree Lynn Rybak, Ph.D. on May 12, 2004 and May 13, 2004. During this interview, the 35 U.S.C. §§112 and 103(a) rejections were discussed, as were declarations needed to overcome the rejections.

Applicants' representative explained that the invention is directed to transgenic plants that express dermaseptin peptides. In addition, Applicants' representative asserted that the claims should not be limited to particular dermaseptin peptides, because such peptides are well known in the art. Evidence for this position on page 7 of the application was noted to the Examiner. Applicants agreed to submit a §1.132 Declaration signed by Dr. Hancock, an expert in the field of antimicrobial peptides, explaining that those in the relevant art understand what a dermaseptin cationic peptide is. The Examiner noted that if the §112 rejections were overcome, the claims could be amended to rejoin the other dermaseptin molecules previously restricted out of the application (namely, SEQ ID NOS: 4-14).

In addition, Applicants' representative explained that the present invention was not obvious to the inventors because of the failures of others to obtain disease resistance in plants transfected with a cationic peptide. Applicants agreed to submit a §1.132 Declaration signed by co-inventor Dr. Misra, explaining why the invention was not obvious to her.

Applicants also agreed to cancel claim 1, and amend claim 2 to clarify that the plant has disease resistance.

Amendments to the Specification

A new abstract on a separate sheet is enclosed.

The specification has been amended to remove hyperlinks, remove redundancies, and to correct obvious typographical errors.

In view of these amendments, Applicants request that the objections to the specification be withdrawn.

Sequence Rules

The specification has been amended to include sequence identifiers for all amino acid sequences of four or more residues. In addition, a new sequence listing is enclosed that now includes SEQ ID NO: 42 (which was previously presented on page 4, line 27 of the specification) .

In view of these amendments, Applicants request that the objections to the specification and sequence listing be withdrawn.

35 U.S.C. §112, second paragraph

Claims 1-9 and 19-20 were rejected under 35 U.S.C. §112, second paragraph on the ground that the phrase “a dermaseptin cationic peptide” does not teach the metes and bounds of the claimed invention. Applicants respectfully disagree and request reconsideration.

The phrase “a dermaseptin cationic peptide” teaches the metes and bounds of the claimed invention, because the specification provides a clear definition of dermaseptins, and those skilled in the art of antimicrobial peptides understand what the term “dermaseptin” means. The specification provides a clear explanation as to what is meant by the phrase “a dermaseptin cationic peptide” starting on page 6 line 29. Furthermore, numerous dermaseptin sequences are known, and are provided in the sequence listing (SEQ ID NOS: 3-14). As stated in the Declaration of Dr. Hancock, a non-inventor who is considered an expert in the art of antimicrobial peptides, those in the art understand that dermaseptin cationic peptides are a family of antimicrobial peptides originally isolated from arboreal frogs that are about 27-34 amino acids in length. Although dermaseptin was first isolated from *Phyllomedusa*, dermaseptins have been isolated from other organisms, such as *Pachymedusa* and *Agalychnis* (see page 7, lines 3-21). New members of the dermaseptin family are identified based on their sequence similarity to known dermaseptin sequences.

In view of the dermaseptin definition and sequences provided in the specification, and the statements made in Dr. Hancock’s §1.132 declaration, Applicants request that the 35 U.S.C. §112, second paragraph rejection be withdrawn.

35 U.S.C. §112, first paragraph

Claims 1-9 and 19-20 were rejected under 35 U.S.C. §112, first paragraph on the ground that the claims do not comply with the written description requirement and as not enabled. Applicants respectfully disagree and request reconsideration.

Sufficient written description for dermaseptin sequences is provided in the specification, and in the sequence listing. The specification, sequence listing, and the knowledge in the art, reasonably convey to one skilled in the antimicrobial peptide art that the inventors had possession of the claimed invention. The term “dermaseptin” as used throughout the specification is understood by those skilled in the art (see §1.132 Declaration signed by Dr. Hancock). As discussed above, several dermaseptin nucleic acid and protein sequences are known. In addition, new dermaseptin sequences are being identified based on their sequence similarity to known dermaseptin sequences. Applicants are not claiming dermaseptin sequences per se, but rather are claiming plants incorporating such

sequences to confer disease resistance. Since the inventors have generated transgenic plants that express dermaseptin peptides which confer disease resistance to plants, and have provided 12 different dermaseptin peptides (SEQ ID NOS: 3-14) with varying amounts of sequence identity to one another, the claims comply with the written description requirement.

The specification as written is enabled for microbial-resistant transgenic plants comprising dermaseptin molecules. For example, page 7, lines 3-21 teaches that known dermaseptin molecules have been isolated from several different organisms. In addition, the sequence listing provides 12 different dermaseptin peptides (SEQ ID NOS: 3-14). The specification also teaches how to make vectors containing dermaseptin, how to use such vectors to make transgenic plants, and how to screen transgenic plants for resistance to disease (see pages 29-34). In addition, such molecular biology techniques are well known in the art. Therefore, since the Applicants have enabled at least 12 different dermaseptin peptides with varying amounts of sequence identity to one another, and methods of making transgenic plants are well known in the art, the specification provides sufficient enablement for claims directed to microbial-resistant transgenic plants comprising dermaseptin.

Therefore, the claims satisfy the written description and enablement requirements, and the 35 U.S.C. §112, first paragraph rejections should be withdrawn.

35 U.S.C. §103(a)

Claims 1-6, 8, 9, 12, 14, and 16-20 were rejected under 35 U.S.C. §103(a) as obvious in view of Scheffler *et al.* (EP 0552 559 A2) and Strahilevitz *et al.* (*Biochem.* 33:10951-60, 1994) and Steinberg *et al.* (U.S. Patent No. 6,025,326). Applicants respectfully disagree and request reconsideration.

Enclosed is a § 1.132 Declaration signed by Dr. Misra stating that use of dermaseptin to confer broad disease resistance to plants was not obvious, due to the teachings at the time the invention was made. For example, as noted on page 8 of the present Office action, Florack *et al.* (*Transgenic Res.* 4:132-41, 1995) teach that expression of a cecropin B peptide did not lead to the predicted resistance in plants. Instead, the cationic peptide was rapidly degraded. Similarly, Hightower *et al.* (*Plant Cell Rep.* 13:295-9, 1994) were unable to confer disease resistance to tobacco plants transformed with cecropin B. In addition, Pang *et al.* (*Gene* 116:165-72, 1992) observed that scorpion insectotoxin was not properly processed in tobacco plants, and did not provide the plants with additional disease protection. Based on these teachings, those working in the art (including the

present inventors) did not expect that cationic peptides (such as dermaseptin) could be expressed in plants at levels that would confer disease resistance.

Additionally, based on the teachings at the time the invention was made, it was not obvious that dermaseptin would work against a broad spectrum of plant pathogens. For example, although Scheffler *et al.* (EP 0 552 559) disclose transgenic plants that include magainin, this application only discloses that magainin provides disease resistance to particular bacteria. That cationic peptides could be used to provide protection against fungi was not disclosed or suggested. In contrast, the inventors have found that dermaseptin can be expressed to provide protection against a large variety of pathogens, such as bacteria, fungi (including *Phytophthora infestans* (late blight)) and viruses. In addition, the present inventors have found that such disease protection can be achieved while expressing low-levels of dermaseptin that are not toxic to the plant.

It was not obvious that dermaseptin could be used to confer disease resistance to plants, and the 35 U.S.C. §103(a) rejection should be withdrawn.

If any matters remain before a Notice of Allowance is issued, the Examiner is invited to telephone the undersigned.

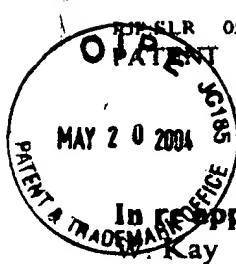
Respectfully submitted,

KLARQUIST SPARKMAN, LLP

By


Sheree Lynn Rybak, Ph.D.
Registration No. 47,913

One World Trade Center, Suite 1600
121 S.W. Salmon Street
Portland, Oregon 97204
Telephone: (503) 226-7391
Facsimile: (503) 228-9446



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: Santosh Misra and William
M. Kay

Application No. 09/936,885

Filed: September 17, 2001

Confirmation No. 2898

For: TRANSGENIC PLANTS THAT ARE
RESISTANT TO A BROAD SPECTRUM
OF PATHOGENS

Examiner: David H. Kruse

Art Unit: 1638

Attorney Reference No. 7013-60993-01

DECLARATION UNDER § 1.132

1. I, Santosh Misra, Ph.D., am a co-inventor named in the above-referenced patent application.
2. I have read and understand the above-referenced patent application, including the pending claims, and the Office action dated February 17, 2004.
3. It is my understanding that in the Office action of February 17, 2004, claims 1-6, 8, 9, 12, 14, and 16-20 were rejected under 35 U.S.C. §103(a) as obvious in view of Scheffler *et al.* (EP 0552 559 A2) and Strahilevitz *et al.* (*Biochem.* 33:10951-60, 1994) and Steinberg *et al.* (U.S. Patent No. 6,025,326). However, it was not obvious to me and my co-inventor Dr. William Kay, that dermaseptin peptides could be used to confer broad disease resistance to plants.
4. At the time the invention was made, there was a significant amount of failure by others in the field. Although several different groups proposed expressing cationic peptides in plants to confer microbial resistance, many were not successful. For example, many journal articles published at the time of the invention disclosed that expression of cationic peptides in plants did not confer microbial resistance. Florack *et al.* (*Transgenic Res.* 4:132-41, 1995) noted that expression cecropin B did not lead to the predicted disease resistance in plants. Instead, the

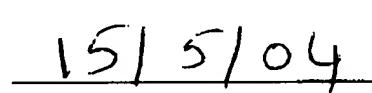
cecropin peptide was rapidly degraded in the plant. Similarly, Hightower *et al.* (*Plant Cell Rep.* 13:295-9, 1994) were unable to confer disease resistance to tobacco plants transformed with cecropin B. Studies using the scorpion insectotoxin indicated that the peptide was not properly processed in tobacco plants, and as a result did not provide the plants with additional disease protection (Pang *et al.*, *Gene* 116:165-72, 1992). Based on these teachings, I did not expect that dermaseptin could be expressed in plants at levels that would confer disease resistance. In addition, there was concern that the amount of dermaseptin that would needed to confer disease resistance would be toxic to the plants. Surprisingly, we found that disease protection was achieved while expressing non-toxic-levels of dermaseptin.

5. It was also not obvious that dermaseptin would be effective against a broad spectrum of plant pathogens. Although some groups were able to confer bacterial resistance by using magainin, that a single cationic peptide could be used to confer resistance to many microbes including yeast, fungi, and bacteria was not obvious. For example, Scheffler *et al.* (EP 0 552 559) only discloses that magainin provides disease resistance to particular bacteria. That cationic peptides could be used to provide protection against fungi was not specifically disclosed or suggested. Surprisingly, we have found that low-levels of dermaseptin can be expressed in plants to confer resistance to bacteria, fungi, and viruses.

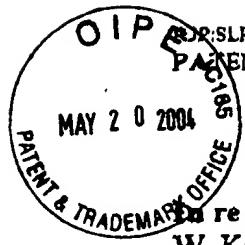
6. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. § 1001, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.



Santosh Misra, Ph.D.



Date



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Da re application of: Santosh Misra and William
W. Kay

Application No. 09/936,885

Filed: September 17, 2001

Confirmation No. 2898

For: TRANSGENIC PLANTS THAT ARE
RESISTANT TO A BROAD SPECTRUM
OF PATHOGENS

Examiner: David H. Kruse

Art Unit: 1638

Attorney Reference No. 7013-60993-01

DECLARATION UNDER 1.132

1. I, Robert E.W. Hancock, Ph.D., am an expert in the field of antimicrobial peptides. I hold a Ph.D. from Adelaide. I am currently a Professor in the Department of Microbiology and Immunology at the University of British Columbia (UBC) in Vancouver, British Columbia, Canada, the Director, Centre for Microbial Diseases and Immunity Research at UBC, and the Canada Research Chair in Microbiology/Genomics and Health cluster at UBC. I presently hold 16 issued patents in the field of cationic peptides, and have over 300 publications. A copy of my Curriculum Vitae is attached (Exhibit A).

2. It is my understanding that some of the claims of the above-referenced patent application were rejected as indefinite, on the ground that the term "dermaseptin cationic peptide" is unclear. It is also my understanding that the position of the United States Patent and Trademark Office (PTO) is that those skilled in the art, such as myself, would not know what the term "dermaseptin cationic peptide" referred to.

3. As one skilled in the art of cationic peptides, I declare that those skilled in the art understand what is meant by the term "dermaseptin cationic peptide." Dermaseptin cationic peptides are a family of antimicrobial peptides originally isolated from arboreal frogs that are about 27-34 amino acids in length. Dermaseptins share properties with other short alpha-helical peptides in their ability to be water soluble and interact with phospholipid membranes. However, dermaseptin peptides are distinguished from other alpha-helical cationic peptides (such as magainins and cecropins) both structurally and functionally.

Those skilled in the art of antimicrobial cationic peptides infer structure and function based on the similarities of the sequences.

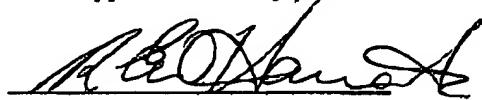
4. Dermaseptin peptides are structurally and functionally different from other cationic peptides. For example, the cecropins, magainins, and dermaseptins vary considerably in chain length, hydrophobicity, and overall distribution of charges. These structural differences lead to functional differences between members of the class of alpha-helical cationic peptides. In addition, because there is no significant sequence homology between dermaseptins and other members of the alpha-helical cationic peptide family, the structure and function of dermaseptins differs. In fact, sequence alignments of dermaseptin molecules with other alpha-helical cationic peptides reveals very low identity. For example, when the dermaseptin amino acid sequence DVLKKIGTVLHAGKAALGAVADTISQ from *Phyllomedusa bicolor* is aligned to the magainin 1 amino acid sequence GIGKFLHSAGKFGKAFVGEIMKS from *X. laevis* only three consecutive amino acids align. Similarly, when the dermaseptin amino acid sequence DVLKKIGTVLHAGKAALGAVADTISQ from *Phyllomedusa bicolor* is aligned to the cecropin amino acid sequence GWLKKIGKKIERVGQHTRDATIQTTLAVAQQAANVAATARG from *Musca domestica* only five consecutive amino acids align. Due to this low level of sequence alignment, those in the art recognize that the structure and function of dermaseptin is distinguishable from those of magainin or cecropin.

5. Those skilled in the art would conclude that a sequence that shared a high amount of sequence alignment with a known dermaseptin sequence would be a member of the dermaseptin family. For example, 22 of the amino acids from the dermaseptin amino acid sequence DVLKKIGTVLHAGKAALGAVADTISQ from *Phyllomedusa bicolor* align with the dermaseptin amino acid sequence TMLKKLGTMALHAGKAALGAAADTISQGTQ from *Phyllomedusa sauvagei*. In addition, even the amino acids that do not align are conservative substitutions. Due to this high level of sequence alignment, those in the art can identify other dermaseptin molecules, and distinguish them from other alpha-helical cationic peptides.

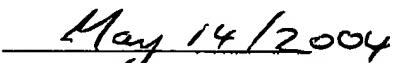
6. The functions of members of the family of cationic peptides include the ability to kill gram positive bacteria, gram negative bacteria, fungi, viruses, nematoes, helminths, and cancer cells. However, each family member has its own unique combination of abilities, although some functions are shared with other family members. For example, dermaseptins have a broader spectrum of antimicrobial abilities than other antimicrobial peptides. Dermaseptin irreversibly inhibits growth of pathogenic fungi, and can also inhibit the growth of bacteria, yeast, viruses, and protozoa. Another difference between dermaseptins and

other cationic peptides is their inability to lyse erythrocytes. Melittin, pardaxin, magainin, and cecropins all have the ability to lyse erythrocytes.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. § 1001, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.



Robert E.W. Hancock, Ph.D.



Date



Curriculum Vitae

Robert Ernest William HANCOCK, OC, FRSC, FAAM, PhD

EMPLOYMENT: Professor, Microbiology & Immunology, University of British Columbia; Director, Centre for Microbial Diseases and Immunity Research (CMDR); Canada Research Chair in Microbiology

EDUCATION: B.Sc. (Hons) Microbiology, University of Adelaide, 1971; Ph.D. Microbiology, University of Adelaide, 1975

APPOINTMENTS

1975-77 Alexander von Humboldt Stipendiat, University Tübingen, Germany
1977-78 Research Bacteriologist I, University of California, Berkeley
1978-83 Assistant Professor, Microbiology, University of British Columbia
1983-86 Associate Professor with tenure, Microbiology, University of British Columbia
1983- Associate Member, Pediatrics, University of British Columbia
1984-85 Vice President, North West Branch, American Society for Microbiology
1985-86 President, North West Branch, American Society for Microbiology
1986- Professor, Microbiology, University of British Columbia
1987-93 Medical Scientific Advisory Cte, Canadian Cystic Fibrosis Foundation, Chair, 90-93.
1989-96 Founding Scientific Director, Canadian Bacterial Diseases Network
1994 WHO Committee on Antimicrobial Resistance and Surveillance
1997- Director, UBC Centre for Microbial Diseases and Host Defence Research
1998-2002 Chair, PseudoCAP (*Pseudomonas aeruginosa* community genome annotation project)
2000 MRC CIHR Institute Simulation committee.
2000 Genome BC implementation committee.
2001-8 Canada Research Chair in Microbiology/Genomics and Health cluster at UBC.
2001 Officer of the Order of Canada
2001-2 President and Co-Founder, Inimex Pharmaceuticals Inc
2002 Fellow, American Academy of Microbiology
2002-5 Co-Director, Functional Pathogenomics of Mucosal Immunity Program

AWARDS AND DISTINCTIONS

Major

- Zellers Senior Scientist award of the Canadian Cystic Fibrosis Foundation, 2004.
- Aventis Pharmaceuticals Award, 2003 (World's top prize for antimicrobial research from American Society of Microbiology).
- Fellow of the American Academy of Microbiology, 2002.
- Jubilee Medal (Commemorative Medal for 50th anniversary of Queen Elizabeth II's reign), 2002.
- Officer of the Order of Canada, 2001. Innovation and Achievement Award, BC Biotech, 2001.
- Canada Research Chair in Microbiology/Genomics and Health, 2001-2008.
- Jacob Biely Faculty Research Prize, 2000 (Leading research prize at UBC).
- Medical Research Council of Canada/CIHR Distinguished Scientist Award, 1995-00.
- Fellow of the Royal Society of Canada, 1994.
- 125th Anniversary of Canada Silver Medal, 1993 for service (to Canadian Cystic Fibrosis Fdn).
- Canadian Society of Microbiologists/New England Biolabs Lecturer, 1992.
- Founding Scientific Director, Canadian Bacterial Diseases Network of Centres of Excellence, 1989-96.
- Canadian Society of Microbiologists Award, 1987 (For outstanding contributions).

EXHIBIT

A

tabbed

Others

- Interview article about career to date in *Lancet Infectious Diseases* 3: 736-739; 2003, entitled "Robert E W Hancock – boosting innate immunity to combat infection"
- Listed on ISI Highly Cited Authors in Microbiology at hcr3.isiknowledge.com/home.cgi, ISI author publication number A1065-2002-Z; 20 papers are cited 111 to 569 times.
- Honorary Member, International Golden Key Society, 2002-3.
- UBC Excellence in Research/ Vancouver Institute Lecture, March 9, 2002.
- Featured interview on HMS Beagle, 2000, at www.biomednet.com/hmsbeagle/71/notes/biofeed.
- UBC Science Undergraduate Society Teaching Excellence Award, 1999-2000.
- Featured in Medical Research Council Performance Report to Parliament, March 1999, pp. 19-20: "International calibre health research. Progress in the search for more effective antibiotics".
- Featured in Lifelines, *Lancet*, May 29, 1998
- UBC Faculty of Science Lecturer, 1992.
- Canadian Who's Who, 1990-04.
- UBC Killam Research Prize, 1988.
- UBC Izaak Walton Killam Memorial Senior Fellowship, 1986-87.
- Foundation for Microbiology lecturer of the American Society for Microbiology, 1985-86.
- Alexander von Humboldt Scholarship for research in West Germany, 1975-77.

SCHOLARLY AND PROFESSIONAL ACTIVITIES

Current Research interests:

Cationic Antimicrobial (Host Defence) Peptides: Mechanism of Action, Structure:Function relationships; Peptide: membrane interactions; Structure of antimicrobial peptides; Aerosol delivery of peptides as a potential therapeutic strategy for cystic fibrosis; Use in transgenic plants.

Cationic Host Defence Peptides: Involvement of peptides in innate immunity in humans, and mice; Microarray studies of peptide interaction with human, mouse, fungal and bacterial cells; Mechanism of interaction of peptides with human epithelial, monocyte and macrophage-like cells; Anti-endotoxic activity of antimicrobial peptides.

Biotechnology: Rational antimicrobial peptide design for improved uptake; Recombinant synthesis of peptide antibiotics; Use of peptides in food preservation; Design of peptides for boosting Innate Immunity and overcoming harmful inflammation.

Outer membranes of Gram negative bacteria: especially *Pseudomonas aeruginosa*. Molecular genetics of outer membrane proteins; outer membrane proteins and porins of other gram negative bacteria; Interactions of outer membrane proteins with host cells, role of the outer membrane in pathogenesis.

Antibiotics: Antibiotics and outer membrane permeability; mechanisms of antibiotic resistance and efflux; Adaptive and mutational resistance to antibiotics

Genomic studies of Pseudomonas aeruginosa: Pathogenomics, functional genomics and informatics; Microarray studies; Construction of *lux*-fusion knockout libraries.

Research or equivalent grants

Research Granting Agencies: Canadian Bacterial Diseases Network; Medical Research Council of Canada (now CIHR); Networks of Centres of Excellence; Canadian Cystic Fibrosis Foundation; CCFF SPARx Program, US CF Foundation, NSERC, Genome Canada.

Research Grant: Funding (Individual totals) 1978-2004, \$15,809,165. Co-funded \$306,660.

Group grants: (As principal applicant and program leader) \$61,031,414.

Equipment Grants: Funding Totals 1978-2004, \$1,192,379. In addition, I participated as an applicant in several joint equipment grants to CFI (totalling more than \$180,000,000).

Research Contracts: Funding Totals 1982-2004, \$1,330,193.

Invited Major Meeting Presentations

(Last 2 years of total 147; More than 200 talks at Universities and Companies not included)

1. UBC Excellence in Research/ Vancouver Institute Lecture, Vancouver, March 9, 2002.
2. BC Biotech Alliance Breakfast lecture, Vancouver, March 13, 2002.
3. CBDN/CMCI AGM 2002, Saskatoon, Canada, June 20-22, 2002.
4. AstraZeneca Symposium on Bacterial Permeability and Efflux, Boston, MA, Nov.8, 2002
5. Biofuture 2002, Vancouver, BC, Nov.21-22, 2002. Genomics session Chair & speaker.
6. Consumer Specialty Products Assn, Annual Meeting, Ft. Lauderdale, Dec.10, 2002.
7. 3rd ASM & TIGR Conference on Microbial Genomes, New Orleans, LA, Jan 29-Feb 1, 2003.
8. Genome BC Genomics Forum, Vancouver, March 27th, 2003.
9. 4th GRC on Antimicrobial Peptides, Barga, Italy, April 27-30, 2003.
10. Pore forming toxins and Maxi-channels. GRTM 21st Symposium, Montreal, May 26-28, 2003.
11. Bovine Genomics Workshop, Montreal, Quebec, June 17-19, 2003.
12. Bio2003 Annual Convention, Washington Convention Center, DC, June 22-25, 2003
13. 2003 International Pseudomonas Mtg, Keynote speaker, Quebec City, September 8, 2003.
14. ICAAC. Aventis Award Lecture, Chicago September 15, 2003.
15. Ann Mtg of Austrian Soc Biochemistry & Molecular Biology, Graz, Austria, Sept. 21-25, 2003.
16. Commercialise 2003. Melbourne, Australia, Nov 18, 2003.
17. Building Biotech Symp., UBC Student Biotechnology Network and BC Biotech, Feb. 26, 2004.
18. Genomics and the Science of Life Forum, Genome BC, March 26, 2004.
19. FDA Proteins and Peptides Workshop, U. Maryland, April 26, 2004.

Conference Organizer. (Last 5 years of total 17 meetings organized)

Conference Committee: North American Cystic Fibrosis Meeting, Montreal, October 15-18, 1998

Vice Chair: 2nd Gordon Conference on Antimicrobial Peptides, April 25-30, 1999

Organizing Committee: Wall Antibiotic Resistance Meeting, April 23-25, 1999

Co-Chair: Gordon Conference on Antimicrobial peptides, Ventura, Ca, March 2001

Organizer: meeting re establishing a Canadian Infection and Immunity Inst., Ottawa, Jan 14, 2000

Co-organizer: meeting re establishing a Canadian Food and Water Safety Network, June 2-4, 2000

Co-organizer: Broken Arrow mtg of the Canadian Cystic Fibrosis Foundn, Toronto, Sept. 7-9, 2001

Chair and Co-founder: 1st Gordon Conference on Multi Drug Efflux Systems, 2003.

Graduate Students Graduated

Thalia I. Nicas, Ph.D. (1978-82); Barbara Angus, Ph.D. (1980-86); Lucy Mutharia, Ph.D. (1980-84); R. Keith Poole, Ph.D. (1981-86); Bernadette Loh, M.Sc. (1981-84); Janet Sawyer, M.Sc. (1985-87); Wendy Woodruff, Ph.D. (1983-88); Angus Bell, Ph.D. (1984-89); Janet Kluftinger, Ph.D. (1984-89); Nancy Martin, Ph.D. (1985-92); Catherine Ullstrom, M.Sc. (1986-90); Kevin Piers, Ph.D. (1987-93); Eileen Rawling, Ph.D. (1988-95); Renee Finnen, M.Sc. (1989-91); Michelle Young, M.Sc. (1990-92); Rebecca Wong, Ph.D. (1990-95); Anand Sukhan, Ph.D. (1991-96); Xiaowen Liao, Ph.D. (1991-96); Hongjin Huang, Ph.D. (1991-95); Maurice Exner, Ph.D. (1991-97); Manhong Wu, Ph.D. (1993-98); Matthew McCusker, M.Sc. (1996-98); Agnes Kwasnicka, M.Sc. (1997-99); Monisha Scott, M.Sc. (1996-98); Kandy Wong, Ph.D. (1994-2001); Carol Friedrich, Ph.D. (1995-2001); Aleks Partzykat, Ph.D. (1996-2001); Monisha Scott, Ph.D. (1998-2002); Jim Jo, M.Sc. (1999-2002).

SERVICE TO THE COMMUNITY

Public Relations Experience

- Especially in my duties as the Scientific Director of the Canadian Bacterial Diseases Network and the Director of CMBR, I have talked to numerous reporters in print, radio and television, appeared on many radio programs including CBC local and national (and Quirks and Quarks twice), CKNW, etc, on television on CBC, CTV, BCTV, King TV, French CBC, Discovery.CA, etc. Appeared in print in the Vancouver Sun (including 3 front page articles), Fortune Magazine, Globe and Mail, Province, Omni, MacLean's, Time, Equinox, Ottawa Citizen, Toronto Star, Edmonton Journal, Calgary Herald,

Montreal Gazette, Canadian Biotech News, Winnipeg Free Press, Bioworld, etc.

- As Chair of the MSAC of the Canadian Cystic Fibrosis Foundation was responsible for reporting to parents and patients about research on cystic fibrosis.
- Participated in 2 large-scale TV productions "Plants that heal" on Discovery channel (replayed throughout the world) and "Antibiotic Resistance" on CBC Prime time (replayed 4 times to date).

Memberships on scholarly committees.

- President, Northwest Branch, American Society for Microbiology, 1985-86.
- Vaccine Evaluation Centre, Advisory Board, 1988-92.
- MRC Grants committee for Microbiology and Infectious Diseases 1982-84.
- Canadian Cystic Fibrosis Foundation Grants Committee, 1987-90.
- Member, Alberta Heritage Found for Med Res Scholarship Applications Advisory Cte, 1989-90.
- Canadian Society of Microbiologists, CSM Awards Committee, 1994-7; Chair, 1997.
- Coordinator of the PseudoCAP (*Pseudomonas aeruginosa* genome community annotation) project with 60 International volunteers, 1997-2000; Co-coordinator with Fiona Brinkman since 2000.
- Fellowship Review Cte, Life Sciences Div., Acad. III, Royal Society of Canada, 1998-2001
- Coordinator of a proposal to create a Canadian Institute for Infectious Diseases and Immunity within CIHR, 1999-2000.
- Member, Executive Group of the Multi-Centre Network for Viral Hepatitis, 2000.
- Member, Executive to coordinate Canadian Food Microbiology activities, 2000-2001.
- UBC CIHR transition committee, 2000
- Vancouver Hospitals HSC Microbial Pathogenesis Review Committee, 2000-2.
- Canada Research Chair College of Reviewers, 2000-3
- Panel member/presenter Mike Smith Foundn for Health Research retreat, Vancouver Feb 1/02.
- Michael Smith Foundation for Health Research, Scientific Advisory Board, 2001-3

Selected Editorial Boards.

Editorial Board, Journal of Bacteriology, 1982-90.

Editorial Board, Infection and Immunity, 1986-89.

Editorial Board, Antimicrobial Agents and Chemotherapy, 1987-05.

International Advisory Board, Current Microbiology and Infection, 1996 -

Drug Resistance Updates, 1997-

Editorial Board, Current Opinion in Anti-infective Drugs, 1998-

Reviewer

Review about 80 manuscripts, grants and reviews per year for Nature, Nature Biotechnology, J. Bacteriology, Infection & Immunity, Antimicrobial Agents & Chemotherapy, Peptides, J. Biological Chemistry, J. Peptide Research, J. Membrane Biology, J. Antimicrobial Chemotherapy, FEBS Letters, Proc Natl Acad Sci., Microbiology, Molecular Microbiology, CIHR, NSERC, Canadian Cystic Fibrosis Foundation, US Cystic Fibrosis Foundation etc.

Industrial Experience.

Micrologix Biotech Inc. This TSE-listed company arose from the research of my laboratory. It is capitalized at around \$80,000,000, has more than 50 employees and is in 2 sets of clinical trials, one phase III (recently completed with success in secondary but not primary objectives for prevention of catheter associated infections), and one phase IIb completed for acne.

CBDN. As Scientific Director of CBDN, I participated in the outlicensing of more than 2 dozen technologies and in the formation of 6 companies. I was an early participant, advisor and SAB member in 3 other companies, *Helix Biomedix*, and *Versicor*.

Inimex Pharmaceuticals Inc. President and Co-founder with Brett Finlay

Consultant

Centocor Corporation, Philadelphia, Pennsylvania, 1983-86; Bristol-Myers-Squibb, Ltd., Syracuse, New York, 1984-91; Oncogen, Seattle, WA, 1988-91; Genta Inc., San Diego, 1993-95; Cubist Pharmaceuticals, 1994; Micrologix Biotech Inc, Vancouver, BC, 1994-98; Affymax Inc., Santa Clara, CA., 1995; Gruppo Lepetit (Marion Merrill Dow), Geranzano, Italy, 1995; Vicuron, San Francisco, CA, 1996- (named Versicor from 1996-2001); Canadian Department of National Defence, Ottawa, 1996-7; Proctor-Gamble, Connecticut, 1997; Canadian Inovatech, Abbotsford, BC, 1997-2000; Pathogenesis, Seattle, 1998; Becton Dickinson, Raleigh Durham, 1998; Geltex, Boston, 1998-9; Synphar Inc, Edmonton, 1998-2000; National Research Council, 1999-2000; Smart & Biggar/Astra, 2000; Helix Biomedix, 2001-; Ortho-McNeil Pharmaceuticals, 2001; Symyx, 2002.

Boards

Board of Directors, Micrologix Biotech Inc., Vancouver, 1992-95; Scientific Advisory Board, Chair, Micrologix Biotech Inc., Vancouver, 1995-98; Scientific Advisory Board, Infectious Diseases Biomedical Inc., Vancouver, 1992-95; Board of Directors, Canadian Foundation for Infectious Diseases, 1994-96; Board of Directors, Chair, David Elford Holding Company, 1995-7.

Board of Directors, Novadex Inc., B.C. 1995-96; Scientific Advisory Board, Versicor, San Francisco, 1995-; Advisory Board, Polydex Inc, New York, 1996-8; Scientific Advisory Board, Welichem, Vancouver, 1996-; Chair, Board of Directors, David Elford Forest Management Ltd., 1997-02; Scientific Advisory Board, Helix Biomedix, Seattle, 2000-; President, Inimex Pharmaceuticals, Vancouver, 2001-02; Board of Directors, Inimex Pharmaceuticals, Vancouver, 2001-.

Publications Record

NB. Listed on ISI Highly Cited Authors in Microbiology at hcr3.isiknowledge.com/home.cgi, ISI author publication number A1065-2002-Z; 20 papers are cited 111 to 569 times.

Robert Ernest William HANCOCK

Date:May/2004

REFEREED JOURNAL PUBLICATIONS

1. Skurray, R.A., R.E.W. Hancock, and P. Reeves. 1974. Con mutants: class of mutants in *Escherichia coli* K-12 lacking a major cell wall protein and defective in conjugation and adsorption of a bacteriophage. *J. Bacteriol.* 119:726-735.
2. Hancock, R.E.W., and P. Reeves. 1975. Bacteriophage resistance in *Escherichia coli* K-12: General pattern of resistance. *J. Bacteriol.* 121:983-993.
3. Hancock, R.E.W., J.K. Davies, and P. Reeves. 1976. Cross resistance between bacteriophages and colicins in *Escherichia coli* K-12. *J. Bacteriol.* 126:1347-1350.
4. Hancock, R.E.W. and P. Reeves. 1976. Lipopolysaccharide-deficient, bacteriophage-resistant mutants of *Escherichia coli* K-12. *J. Bacteriol.* 127:98-108.
5. Hancock, R.E.W., and V. Braun. 1976. Nature of the energy requirement for the irreversible adsorption of bacteriophages T1 and ϕ 80 to *Escherichia coli*. *J. Bacteriol.* 12:409-415.
6. Hancock, R.E.W., and V. Braun. 1976. The colicin I receptor of *Escherichia coli* K-12 has a role in enterochelin-mediated iron transport. *FEBS Lett.* 65:208-210.
7. Braun, V., R.E.W. Hancock, K. Hantke, and A. Hartmann. 1976. Functional organization of the outer membrane of *Escherichia coli*. Phage and colicin receptors as components of iron uptake systems. *J. Supramolecular Structure* 5:37-58.
8. Hancock, R.E.W., K. Hantke, and V. Braun. 1976. Iron transport in *Escherichia coli* K-12: Involvement of the colicin B receptor and of a citrate-inducible protein. *J. Bacteriol.* 127:1370-1375.
9. Hancock, R.E.W., K. Hantke, and V. Braun. 1977. Iron transport in *Escherichia coli* K-12: 2,3-dihydroxybenzoate-promoted iron uptake. *Arch. Microbiol.* 114:231-239.
10. Hancock, R.E.W., and H. Nikaido. 1978. Outer membrane of Gram negative bacteria. XIX Isolation from *Pseudomonas aeruginosa* PAO1 and use in reconstitution and definition of the permeability barrier. *J. Bacteriol.* 136:381-390.

11. Hancock, R.E.W., and G.M. Decad and H. Nikaido. 1979. Identification of the protein producing transmembrane diffusion pores in the outer membrane of *Pseudomonas aeruginosa* PA01. *Biochim. Biophys. Acta.* 554:323-331.
12. Hancock, R.E.W., and A.M. Carey. 1979. Outer membrane of *Pseudomonas aeruginosa*. Heat-and 2-mercaptoethanol-modifiable proteins. *J. Bacteriol.* 140:902-910.
13. Chen, U., R.E.W. Hancock, and R. Mishell. 1980. Mitogenic effects of purified outer membrane proteins from *Pseudomonas aeruginosa*. *Infect. Immun.* 28:178-184.
14. Hancock, R.E.W., and A.M. Carey. 1980. Protein D1 - a glucose-inducible, pore-forming protein from the outer membrane of *Pseudomonas aeruginosa*. *FEMS Microbiol. Letters* 8:105-109.
15. Nicas, T.I., and R.E.W. Hancock. 1980. Outer membrane protein H1 of *Pseudomonas aeruginosa*: Involvement in adaptive and mutational resistance to ethylenediamine tetraacetate, polymyxin B and gentamicin. *J. Bacteriol.* 143:872-878.
16. Hancock, R.E.W., R.T. Irvin, J.W. Costerton, and A.M. Carey. 1981. *Pseudomonas aeruginosa* outer membrane: peptidoglycan associated proteins. *J. Bacteriol.* 145:628-631.
17. Hancock, R.E.W., V.J. Raffle, and T.I. Nicas. 1981. Involvement of the outer membrane in gentamicin and streptomycin uptake and killing in *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 19:777-785.
18. Benz, R., and R.E.W. Hancock. 1981. Properties of the large ion-permeable pores formed from protein F of *Pseudomonas aeruginosa* in lipid bilayer membranes. *Biochim. Biophys. Acta* 646:298-308.
19. Angus, B.L., A.M. Carey, D.A. Caron, A.M.B. Kropinski, and R.E.W. Hancock. 1982. Outer membrane permeability in *Pseudomonas aeruginosa*: Comparison of a wild-type with an antibiotic-supersusceptible mutant. *Antimicrob. Agents Chemother.* 21:299-309.
20. Kropinski, A.M.B., J. Kuzio, B.L. Angus, and R.E.W. Hancock. 1982. Chemical and chromatographic analysis of lipopolysaccharide from an antibiotic-supersusceptible mutant of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 21:310-319.
21. Hancock, R.E.W., K. Poole, and R. Benz. 1982. Outer membrane protein P of *Pseudomonas aeruginosa*: Regulation by phosphate deficiency and formation of small anion-specific channels in lipid bilayer membranes. *J. Bacteriol.* 150:730-38.
22. Hancock, R.E.W., A.A. Wieczorek, L.M. Mutharia, and K. Poole. 1982. Monoclonal antibodies against *Pseudomonas aeruginosa* outer membrane antigens: Isolation and characterization. *Infect. Immun.* 37:166-171.
23. Mutharia, L.M., T. I. Nicas, and R.E.W. Hancock. 1982. Outer membrane proteins of *Pseudomonas aeruginosa* serotype strains. *J. Infect. Dis.* 146:770-779.
24. Downum, K.R., R.E.W. Hancock, and G.H.N. Towers. 1982. Mode of action of alpha-terthienyl on *Escherichia coli*. Evidence for a photodynamic effect on membranes. *Photochem. Photobiol.* 36:517-523.
25. Nakajima, K., K. Muroga, and R.E.W. Hancock. 1983. Comparison of fatty acid, protein and serological properties distinguishing outer membranes of *Pseudomonas anguilliseptica* strains from those of fish pathogens and other *Pseudomonads*. *Int. J. Systemat. Bacteriol.* 33:1-8.
26. Nicas, T.I., and R.E.W. Hancock. 1983. Alteration of susceptibility to EDTA, polymyxin B and gentamicin in *Pseudomonas aeruginosa* by divalent cation regulation of outer membrane protein H1. *J. Gen. Microbiol.* 129:509-517.
27. Poole, K., and R.E.W. Hancock. 1983. Secretion of alkaline phosphatase and phospholipase C in *Pseudomonas aeruginosa* is specific and does not involve an increase in outer membrane permeability. *FEMS Microbiol. Lett.* 16:25-29.
28. Nicas, T.I., and R.E.W. Hancock. 1983. *Pseudomonas aeruginosa* outer membrane permeability: Isolation of a porin protein F-deficient mutant. *J. Bacteriol.* 153:281-285.
29. Darveau, R.P., and R.E.W. Hancock. 1983. Procedure for isolation of bacterial lipopolysaccharides from both smooth and rough *Pseudomonas aeruginosa* and *Salmonella typhimurium* strains. *J. Bacteriol.* 155:831-838.
30. Benz, R., M. Gimple, K. Poole and R.E.W. Hancock. 1983. An anion-selective channel from the *Pseudomonas aeruginosa* outer membrane. *Biochim. Biophys. Acta* 730:387-390.
31. Angus, B.L., and R.E.W. Hancock. 1983. Outer membrane porin proteins F, P and D1 of *Pseudomonas aeruginosa* and Pho E of *Escherichia coli* : Chemical crosslinking to reveal native oligomers. *J. Bacteriol.* 155:1042-1051.
32. Mutharia, L.M., and R.E.W. Hancock. 1983. Surface localization of *Pseudomonas aeruginosa*

outer membrane porin protein F by using monoclonal antibodies. *Infect. Immun.* 42:1027-1033.

33. Hancock, R.E.W., L.M. Mutharia, L. Chan, R.P. Darveau, D.P. Speert, and G.B. Pier. 1983. *Pseudomonas aeruginosa* isolates from patients with cystic fibrosis: a class of serum-sensitive, nontypable strains deficient in lipopolysaccharide O side-chains. *Infect. Immun.* 42:170-177.

34. Hancock, R.E.W., K. Poole, M. Gimple and R. Benz. 1983. Modification of the conductance, selectivity and concentration-dependent saturation of *Pseudomonas aeruginosa* protein P channels by chemical acetylation. *Biochim. Biophys. Acta*, 735:137-144.

35. Whitfield, C., R.E.W. Hancock, and J.W. Costerton. 1983. Outer membrane protein K of *Escherichia coli*: Purification and pore-forming properties in lipid bilayer membranes. *J. Bacteriol.* 156:873-879.

36. Darveau, R.P., S. MacIntyre, J.T. Buckley, and R.E.W. Hancock. 1983. Purification and reconstitution in lipid bilayer membranes of an outer membrane pore-forming protein of *Aeromonas salmonicida*. *J. Bacteriol.* 156:1006-1011.

37. Darveau, R.P., W.T. Charnetzky, R.E. Hurlbert, and R.E.W. Hancock. 1983. Effects of growth temperature, 47-megadalton plasmid, and calcium deficiency on the outer membrane protein porin and lipopolysaccharide composition of *Yersina pestis* EV76. *Infect. Immun.* 42:1092-1101.

38. Lam, J.S., L.M. Mutharia, R.E.W. Hancock, N. Hoiby, K. Lam, L. Baek, and J.W. Costerton. 1983. Immunogenicity of *Pseudomonas aeruginosa* outer membrane antigens examined by crossed immunoelectrophoresis. *Infect. Immun.* 42:88-98.

39. Downum, K.R., R.E.W. Hancock and G.H.N. Towers. 1983. Photodynamic action on *Escherichia coli* of natural acetylenic thiophenes, particularly 5-(buten-1-ynyl)-2,2'-bithienyl. *Photobiochem. Photobiophys.* 6:145-152.

40. Benz, R., K. Poole, and R.E.W. Hancock. 1984. Characterization and chemical modification of small anion specific channels formed in lipid bilayer membranes by outer membrane protein P of *Pseudomonas aeruginosa*. *Biophys. J.* 45:81-82.

41. Hancock, R.E.W., E. Mouat, and D.P. Speert. 1984. Quantitation and identification of antibodies to the outer-membrane proteins of *Pseudomonas aeruginosa* in sera of patients with cystic fibrosis. *J. Infect. Dis.* 149:220-226.

42. Benz, R., R.P. Darveau, and R.E.W. Hancock. 1984. Outer-membrane protein PhoE from *Escherichia coli* forms anion-selective pores in lipid-bilayer membranes. *Eur. J. Biochem.* 140:319-324.

43. Darveau, R.P., R.E.W. Hancock and R. Benz. 1984. Chemical modification of the anion selectivity of the PhoE porin from the *Escherichia coli* outer membrane. *Biochim. Biophys. Acta* 774:69-74.

44. Mutharia, L.M., G. Crockford, W.C. Bogard, and R.E.W. Hancock. 1984. Monoclonal antibodies specific for *Escherichia coli* J-5 lipopolysaccharide: Cross-reaction with other gram-negative bacterial species. *Infect. Immun.* 45:631-636.

45. Loh, B., C. Grant and R.E.W. Hancock. 1984. Use of the fluorescent probe 1-N-phenylnaphthylamine to study the interactions of aminoglycoside antibiotics with the outer membrane of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 26:546-551.

46. Hancock, R.E.W., and P.G.W. Wong. 1984. Compounds which increase the permeability of the *Pseudomonas aeruginosa* outer membrane. *Antimicrob. Agents Chemother.* 26:48-52.

47. Poole, K. and R.E.W. Hancock. 1984. Phosphate transport in *Pseudomonas aeruginosa*: Involvement of a periplasmic phosphate-binding protein. *Eur. J. Biochem.*, 144:607-612.

48. Moore, R.A., L. Chan, and R.E.W. Hancock. 1984. Evidence for two distinct mechanisms of resistance to polymyxin B in *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 26:539-545.

49. Hindahl, M.S., G.W.K. Crockford, and R.E.W. Hancock. 1984. Outer membrane protein NmpC of *Escherichia coli*: Pore-forming properties in black lipid bilayers. *J. Bacteriol.* 159:1053-1055.

50. Mutharia, L.M., and R.E.W. Hancock. 1985. Characterization of two surface-localized antigenic sites on porin protein F of *Pseudomonas aeruginosa*. *Canad. J. Microbiol.* 31:381-386.

51. Benz, R., A. Schmidt, and R.E.W. Hancock. 1985. Ion selectivity of gram-negative bacterial porins. *J. Bacteriol.* 162:722-727.

52. Hancock, R.E.W., L.M. Mutharia and E.C.A. Mouat. 1985. Immunotherapeutic potential of monoclonal antibodies against *Pseudomonas aeruginosa* protein F. *Europ. J. Clin. Microbiol.* 4: 224-227.

53. Mutharia, L.M., and R.E.W. Hancock. 1985. Monoclonal antibody for an outer membrane lipoprotein of the *Pseudomonas fluorescens* group of the family *Pseudomonadaceae*. *Int. J.*

Systemat. Bacteriol. 35:530-532.

54. Peterson, A.A., R.E.W. Hancock, and E.J. McGroarty. 1985. Binding of polycationic antibiotics and polyamines to lipopolysaccharides of *Pseudomonas aeruginosa*. J. Bacteriol. 164: 1256-1261.

55. Comeau, Y., J.J. Hall, R.E.W. Hancock, and W.K. Oldham. 1986. Biochemical model for enhanced biological phosphorous removal. Water Res. 20:1511-1521.

56. Moore, R.A., N.C. Bates and R.E.W. Hancock. 1986. Interaction of polycationic antibiotics with *Pseudomonas aeruginosa* lipopolysaccharide and lipid A studied by using dansyl-polymyxin. Antimicrob. Agents Chemother. 29: 496-500.

57. Parr, T.R., K. Poole, G.W.K. Crockford, and R.E.W. Hancock. 1986. Lipopolysaccharide-free *Escherichia coli* Omp F and *Pseudomonas aeruginosa* protein P porins are functionally active in lipid bilayer membranes. J. Bacteriol. 165: 523-526.

58. Hancock, R.E.W., and R. Benz. 1986. Demonstration and chemical modification of a specific phosphate binding site in the phosphate-starvation-inducible outer membrane porin protein P of *Pseudomonas aeruginosa*. Biochim. Biophys Acta 860: 699-707.

59. Poole, K., and R.E.W. Hancock. 1986. Isolation of a Tn501 insertion mutant lacking porin protein P of *Pseudomonas aeruginosa*. Molec. Gen. Genetics 202:403-409.

60. Poole, K., and R.E.W. Hancock. 1986. Phosphate-starvation-induced outer membrane proteins of the families *Enterobacteriaceae* and *Pseudomonadaceae*: demonstration of immunological cross-reactivity with an antiserum specific for porin protein P of *Pseudomonas aeruginosa*. J. Bacteriol. 165: 987-993.

61. Armstrong, S.K., T.R. Parr, C.D. Parker, and R.E.W. Hancock. 1986. *Bordetella pertussis* major outer membrane porin protein forms small, anion-selective channels in lipid bilayer membranes. J. Bacteriol. 166: 212-216.

62. Woodruff, W.A., T.R. Parr, R.E.W. Hancock, L. Hanne, T.I. Nicas, and B. Iglewski. 1986. Expression in *Escherichia coli* and function of *Pseudomonas aeruginosa* outer membrane porin protein F. J. Bacteriol. 167: 473-479.

63. Hancock, R.E.W., A. Schmidt, K. Bauer, and R. Benz. 1986. Role of lysines in ion selectivity of bacterial outer membrane porins. Biochim. Biophys. Acta 860: 263-267.

64. Huyer, M., T.R. Parr, R.E.W. Hancock and W.J. Page. 1986. Outer membrane porin protein of *Campylobacter jejuni*. FEMS Microbiol Lett. 37: 247-250.

65. Hancock, R.E.W. 1986. Intrinsic antibiotic resistance of *Pseudomonas aeruginosa*. J. Antimicrob. Chemother. 18: 653-659.

66. Moore, R.A., and R.E.W. Hancock. 1986. Involvement of outer membrane of *Pseudomonas cepacia* in aminoglycoside and polymyxin resistance. Antimicrob. Agents Chemother. 30: 923-926.

67. Kropinski, A.M., T.R. Parr, B. Angus, R.E.W. Hancock, W.C. Ghiorse, and E.P. Greenberg. 1987. Isolation of the outer membrane and characterization of the major outer membrane protein of *Spirocheata aurantia*. J. Bacteriol. 169:172-179.

68. Benz, R., and R.E.W. Hancock. 1987. Mechanism of ion transport through the anion-selective channel of the *Pseudomonas aeruginosa* outer membrane. J. Gen. Physiol. 89: 275-295.

69. Poole, K., T.R. Parr and R.E.W. Hancock. 1987. Phosphate-selective porins from the outer membranes of fluorescent *Pseudomonas* sp. Canad. J. Microbiol. 33: 63-69.

70. Parr, T.R., R.A. Moore, L.V. Moore and R.E.W. Hancock. 1987. Role of porins in intrinsic antibiotic resistance of *Pseudomonas cepacia*. Antimicrob. Agents Chemother. 31: 121-123.

71. Bayer, A.S., J. Peters, T.R. Parr, L. Chan and R.E.W. Hancock. 1987. Role of β -lactamase in in vivo development of ceftazidime resistance in experimental *Pseudomonas aeruginosa* endocarditis. Antimicrob. Agents Chemother. 31: 253-258.

72. Hancock, R.E.W. 1987. Role of porins in outer membrane permeability. J. Bacteriol. 169: 929-933.

73. Speert, D.P., J.E. Dimmick, G.B. Pier, J.M. Saunders, R.E.W. Hancock and N.M. Kelly. 1987. An immunohistological evaluation of *Pseudomonas aeruginosa* pulmonary infection in two patients with cystic fibrosis. Pediatr. Res. 22:743-747.

74. Battershill, J., D.P. Speert, and R.E.W. Hancock. 1987. Use of monoclonal antibodies to protein F of *Pseudomonas aeruginosa* as opsonins for phagocytosis by macrophages. Infect. Immun. 55:2531-2533.

75. Kelly, N.M., J.P. Arbuthnott, J. Battershill, S. Kuo, and R.E.W. Hancock. 1987. Colonial dissociation and susceptibility to phagocytosis of *Pseudomonas aeruginosa* grown in a chamber

implant model in mice. *Infect. Immun.*, 55:2841-2843.

76. Angus, B.L., J. Fyfe and R.E.W. Hancock. 1987. The hydrophobic uptake pathway across the outer membrane of the antibiotic supersusceptible *Pseudomonas aeruginosa* mutant Z61. *FEMS Microbiol. Lett.*, 48:153-57.

77. Angus, B.L., J. Fyfe and R.E.W. Hancock. 1987. Mapping and characterization of two mutations to antibiotic supersusceptibility in *Pseudomonas aeruginosa*. *J. Gen. Microbiol.*, 133:2905-2914.

78. Lam, J.S., M.Y.C. Lam, L.A. MacDonald and R.E.W. Hancock. 1987. Visualization of *Pseudomonas aeruginosa* O antigens by using a protein A-dextran-colloidal gold conjugate with both immunoglobulin G and immunoglobulin M in monoclonal antibodies. *J. Bacteriol.*, 169:3531-3538.

79. Sawyer, J.G., N.L. Martin and R.E.W. Hancock. 1988. Interaction of macrophage cationic proteins with the outer membrane of *Pseudomonas aeruginosa*. *Infect. Immun.* 56:693-698.

80. Worobec, B., R. Siehnel, P. Gladman and R.E.W. Hancock. 1988. Gene cloning and expression of the *Pseudomonas aeruginosa* periplasmic phosphate-binding protein. *FEMS Microbiol. Lett.* 52:235-238.

81. Worobec, B., N.L. Martin, W. McCubbin, C. Kay, G. Brayer and R.E.W. Hancock. 1988. Large-scale purification and biochemical characterization of crystallization-grade porin protein P from *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta* 939:366-374.

82. Woodruff, W.A. and R.E.W. Hancock. 1988. Construction and characterization of *Pseudomonas aeruginosa* protein F-deficient mutants after in vitro and in vivo insertion mutagenesis of the cloned gene. *J. Bacteriol.* 170:2592-2598.

83. Siehnel, R., E.A. Worobec, and R.E.W. Hancock. 1988. Cloning of the *Pseudomonas aeruginosa* outer membrane porin protein P gene: Evidence for a linked region of DNA homology. *J. Bacteriol.* 170:2312-2318.

84. Rivera, M., L.E. Bryan, R.E.W. Hancock and E.J. McGroarty. 1988. Heterogeneity of lipopolysaccharides from *Pseudomonas aeruginosa*. Analysis of lipopolysaccharide chain length. *J. Bacteriol.* 170:512-521.

85. Rivera, M., R.E.W. Hancock, J.G. Sawyer, A. Haug and E.J. McGroarty. 1988. Enhanced binding of polycationic antibiotics to lipopolysaccharide from an aminoglycoside-supersusceptible, tolA mutant strain of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 32:649-655.

86. Moon, M.M., L.D. Hazlett, R.E.W. Hancock, R.S. Berk and R. Barrett. 1988. Monoclonal antibodies provide protection against ocular *Pseudomonas aeruginosa* infection. *Invest. Ophthalmol. Vis. Sci.* 29:1277-1284.

87. Siehnel, R.J., E. Worobec, and R.E.W. Hancock. 1988. Regulation of components of the *Pseudomonas aeruginosa* phosphate-starvation-inducible regulon in *Escherichia coli*. *Molec. Microbiol.* 2:347-352.

88. Hancock, R.E.W. and L. Chan. 1988. Outer membranes of environmental isolates of *Pseudomonas aeruginosa*. *J. Clin. Microbiol.* 26:2423-2424.

89. Hancock, R.E.W. and A. Bell. 1988. Antibiotic uptake into Gram-negative bacteria. *Europ. J. Clin. Microbiol. Infect. Dis.* 7:713-720.

90. Kelly, N.M., A. Bell and R.E.W. Hancock. 1989. Surface characteristics of *Pseudomonas aeruginosa* grown in a chamber implant model in mice and rats. *Infect. Immun.* 57:344-350.

91. Kluftinger, J., F. Lutz and R.E.W. Hancock. 1989. *Pseudomonas aeruginosa* cytotoxin: Periplasmic localization and inhibition of macrophages. *Infect. Immun.* 57:882-886.

92. Kluftinger, J.L., N.M. Kelly and R.E.W. Hancock. 1989. Stimulation by fibronectin of macrophage-mediated phagocytosis of *Pseudomonas aeruginosa*. *Infect. Immun.* 57:817-822.

93. Bell, A., and R.E.W. Hancock. 1989. Outer membrane protein H1 of *Pseudomonas aeruginosa*: Purification of the protein and cloning and nucleotide sequence of the gene. *J. Bacteriol.* 171:3211-3217.

94. Woodruff, W.A., and R.E.W. Hancock. 1989. *Pseudomonas aeruginosa* outer membrane protein F: Structural role and relationship to the *Escherichia coli* OmpA protein. *J. Bacteriol.* 171:3304-3309.

95. Lee, H.S., R.E.W. Hancock, and J.L. Ingraham. 1989. Properties of a *Pseudomonas stutzeri* outer membrane channel-forming protein (NosA) required for production of copper-containing N₂O reductase. *J. Bacteriol.* 171:2096-2100.

96. Kelly, N.M., E.G. Rawling and R.E.W. Hancock. 1989. Determinants of the efficacy of tobramycin therapy against isogenic nonmucoid and mucoid derivatives of *Pseudomonas*

aeruginosa PA01 growing in peritoneal chambers in mice. *Antimicrob. Agents Chemother.* 33:1207-1211.

97. Kluftinger, J.L., N.M. Kelly, B.H. Jost and R.E.W. Hancock. 1989. Fibronectin as an enhancer of nonopsonic phagocytosis of *Pseudomonas aeruginosa* by macrophages. *Infect. Immun.* 57:2782-2785.

98. Kelly, N.M., J.L. Kluftinger, B.L. Pasloske, W. Paranchych, and R.E.W. Hancock. 1989. *Pseudomonas aeruginosa* pili as ligands for nonopsonic phagocytosis by fibronectin-stimulated macrophages. *Infect. Immun.* 57:3841-3845.

99. Kelly, N.M., M.H. MacDonald, N. Martin, T.I. Nicas, and R.E.W. Hancock. 1990. Comparison of the outer membrane protein and lipopolysaccharide profiles of mucoid and nonmucoid *Pseudomonas aeruginosa*. *J. Clin. Microbiol.* 28:2017-2021.

100. Siehnel, R.J., N.L. Martin, and R.E.W. Hancock. 1990. Sequence and relatedness in other bacteria of the *Pseudomonas aeruginosa* oprP gene coding for the phosphate-specific porin P. *Molec. Microbiol.* 4:831-838.

101. Hancock, R.E.W., R. Siehnel and N. Martin. 1990. Outer membrane proteins of *Pseudomonas*. *Molec. Microbiol.* 4:1069-1075.

102. Ullstrom, C.A., R. Siehnel, W. Woodruff, S. Steinbach, and R.E.W. Hancock. 1991. Conservation of the gene for outer membrane protein OprF in the family *Pseudomonadaceae*: Sequence of the *Pseudomonas syringae* OprF gene. *J. Bacteriol.* 173:768-775.

103. Hancock, R.E.W., S.W. Farmer, Z. Li, and K. Poole. 1991. Interaction of aminoglycosides with the outer membranes and purified lipopolysaccharide and OmpF porin of *Escherichia coli*. *Antimicrob. Agents Chemother.* 35:1309-1314.

104. Walker, S.G., R.E.W. Hancock, and J. Smit. 1991. Expression in *Caulobacter crescentus* of the phosphate-starvation-inducible porin OprP of *Pseudomonas aeruginosa*. *FEMS Microbiol. Letters* 77:217-222.

105. Bellido, F., J.-C. Pechere, and R.E.W. Hancock. 1991. Novel method for measurement of outer membrane permeability to new β -lactams in intact *Enterobacter cloacae* cells. *Antimicrob. Agents Chemother.* 35:68-72.

106. Bellido, F., J.-C. Pechere, and R.E.W. Hancock. 1991. Reevaluation of the factors involved in the efficacy of new β -lactams against *Enterobacter cloacae*. *Antimicrob. Agents Chemother.* 35:73-78.

107. Saravolac, E.G., N.F. Taylor, R. Benz, and R.E.W. Hancock. 1991. Purification of glucose-inducible outer membrane protein OprB of *Pseudomonas putida* and reconstitution of glucose-specific pores. *J. Bacteriol.* 173:4970-4976.

108. Bell, A., M. Bains, and R.E.W. Hancock. 1991. *Pseudomonas aeruginosa* outer membrane protein OprH: Expression from the cloned gene and function in EDTA and gentamicin resistance. *J. Bacteriol.* 173:6657-6664.

109. Hancock, R.E.W., and F. Bellido. 1992. Factors involved in the enhanced efficacy against Gram-negative bacteria of fourth generation cephalosporins. *J. Antimicrob. Chemother.* 29, Suppl. A:1-6.

110. Hancock, R.E.W., and F. Bellido. 1992. Antibiotic uptake: unusual results for unusual molecules. *J. Antimicrob. Chemother.* 29:235-243.

111. Farmer, S., Z. Li, and R.E.W. Hancock. 1992. Influence of outer membrane mutations on susceptibility of *Escherichia coli* to the dibasic macrolide azithromycin. *J. Antimicrob. Chemother.* 29:27-33.

112. Piddock, L.J.V., M.C. Hall, F. Bellido, M. Bains, and R.E.W. Hancock. 1992. A pleiotropic, post-therapy, enoxacin-resistant mutant of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 36:1057-1061.

113. Hancock, R.E.W., C. Egli, R. Benz, and R.J. Siehnel. 1992. Overexpression in *Escherichia coli* and functional analysis of a novel PP_i selective porin, OprO, from *Pseudomonas aeruginosa*. *J. Bacteriol.* 174:471-476.

114. Siehnel, R.J., C. Egli, and R.E.W. Hancock. 1992. Polyphosphate-selective porin OprO of

Pseudomonas aeruginosa: Expression, purification and sequence. *Molec. Microbiol.* 6:2319-2326.

115. Jaeger, K.E., F.-J. Adrian, H.E. Meyer, R.E.W. Hancock, and U.K. Winkler. 1992. Extracellular lipase from *Pseudomonas aeruginosa* is an amphiphilic protein. *Biochem. Biophys. Acta* 1120:315-321.

116. Karunaratne, D.N., J.C. Richards and R.E.W. Hancock. 1992. Characterization of lipid A from *Pseudomonas aeruginosa* O-antigenic B-band lipopolysaccharide by 1D and 2D NMR and mass spectral analysis. *Arch. Biochem. Biophys.* 299:368-376.

117. Bellido, F., N.L. Martin, R.J. Siehnel, and R.E.W. Hancock. 1992. Reevaluation, using intact cells, of the exclusion limit and role of porin OprF in *Pseudomonas aeruginosa* outer membrane permeability. *J. Bacteriol.* 174:5196-5203.

118. Finnen, R.L., N.L. Martin, R.J. Siehnel, W.A. Woodruff, M. Rosok, and R.E.W. Hancock. 1992. Analysis of the *Pseudomonas aeruginosa* major outer membrane protein OprF by use of truncated OprF derivatives and monoclonal antibodies. *J. Bacteriol.* 174:4977-4985.

119. McCutcheon, A.R., S.M. Ellis, R.E.W. Hancock and G.H.N. Towers. 1992. Antibiotic screening of medicinal plants of the British Columbian native peoples. *J. Ethnopharmacol.* 37:213-223.

120. Kennel, W.L., C. Egli, R.E.W. Hancock, and S.C. Holt. 1992. Pore-forming ability of major outer membrane proteins from *Wolinella recta* ATCC 33238. *Infect. Immun.* 60:380-384.

121. Young, M., and R.E.W. Hancock. 1992. Fluoroquinolone supersusceptibility mediated by outer membrane protein OprH overexpression in *Pseudomonas aeruginosa*: Evidence for involvement of a non-porin pathway. *Antimicrob. Agents Chemother.* 36:2365-2369.

122. Young, M.L., M. Bains, A. Bell and R.E.W. Hancock. 1992. Role of *Pseudomonas aeruginosa* outer membrane protein OprH in polymyxin and gentamicin resistance: Isolation of an OprH-deficient mutant by gene replacement techniques. *Antimicrob. Agents Chemother.* 36:2566-2568.

123. Huang, H., R.J. Siehnel, F. Bellido, E. Rawling, and R.E.W. Hancock. 1992. Analysis of two gene regions involved in the expression of the imipenem-specific, outer membrane porin protein OprD of *Pseudomonas aeruginosa*. *FEMS Microbiology Letters* 97:267-274.

124. Hancock, R.E.W., and S.W. Farmer. 1993. Mechanism of uptake of deglucoteicoplanin amide derivatives across outer membranes of *Escherichia coli* and *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 37:453-456.

125. Benz, R., C. Egli, and R.E.W. Hancock. 1993. Anion transport through the phosphate-specific OprP-channel of the *Pseudomonas aeruginosa* outer membrane: Effects of phosphate, di- and tri-basic anions and of negatively-charged lipids. *Biochim. Biophys. Acta*, 1149: 224-230.

126. Egli, C., W.K. Leung, K.H. Müller, R.E.W. Hancock, and B.C. McBride. 1993. Pore-forming properties of the major 53-kilodalton surface antigen from the outer sheath of *Treponema denticola*. *Infect. Immun.* 61:1694-1699.

127. Mork, T., and R.E.W. Hancock. 1993. Mechanisms of nonopsonic phagocytosis of *Pseudomonas aeruginosa*. *Infect. Immun.* 61: 3287-3293.

128. Piers, K.L., M.H. Brown, and R.E.W. Hancock. 1993. Recombinant DNA procedures for producing small antimicrobial cationic peptides in bacteria. *Gene* 134:7-13.

129. Wong, R.S.Y., H. Jost, and R.E.W. Hancock. 1993. Linker-insertion mutagenesis of *Pseudomonas aeruginosa* outer membrane protein OprF. *Molec. Microbiol.* 10:283-292.

130. Martin, N.M. E. G. Rawling, R. Wong, M. Rosok, and R.E.W. Hancock. 1993. Conservation of surface epitopes in *Pseudomonas aeruginosa* outer membrane porin protein OprF. *FEMS Microbiol. Lett.* 113:261-266.

131. Karunaratne, D.N., S. Farmer and R.E.W. Hancock. 1993. Synthesis of bulky β -lactams for inhibition of cell surface β -lactamase activity. *Bioconjugate Chemistry.* 4:434-439.

132. Huang, H., and R.E.W. Hancock. 1993. Genetic definition of the substrate selectivity of outer membrane porin protein OprD of *Pseudomonas aeruginosa*. *J. Bacteriol.* 175:7793-7800.

133. Piers, K., and R.E.W. Hancock. 1994. The interaction of a recombinant cecropin/melittin hybrid

peptide with the outer membrane of *Pseudomonas aeruginosa*. *Molec. Microbiol.* **12**:951-958.

134. Piers, K.L., M.H. Brown and R.E.W. Hancock. 1994. Improvement of outer membrane-permeabilizing and lipopolysaccharide-binding activities of an antimicrobial cationic peptide by C-terminal modification. *Antimicrob. Agents Chemother.* **38**:2311-2316.

135. McCutcheon, A.R., S.M. Ellis, R.E.W. Hancock and G.H.N. Towers. 1994. Antifungal screening of medicinal plants of British Columbian native peoples. *J. Ethnopharmacol.* **44**:157-169.

136. Blanco, D.R., K. Reimann, J. Skare, C.I. Champion, D. Foley, M.M. Exner, R.E.W. Hancock, J.N. Miller, and M.A. Lovett. 1994. Isolation of the outer membranes from *Treponema pallidum* and *Treponema vincentii*. *J. Bacteriol.* **176**:6088-6099.

137. Saxena, G., A.R. McCutcheon, S. Farmer, G.H.N. Towers and R.E.W. Hancock. 1994. Antimicrobial constituents of *Rhus glabra*. *J. Ethnopharmacol.* **42**:95-99.

138. Saxena, G., S. Farmer, R.E.W. Hancock, and G.H.N. Towers. 1995. Antimicrobial compounds from *Alnus rubra*. *Intl. J. Pharmacognosy* **33**:33-36.

139. Saxena, G., S. Farmer, G.H.N. Towers, and R.E.W. Hancock. 1995. Use of specific dyes in the detection of antimicrobial compounds from crude plant extracts using a thin layer chromatography agar overlay technique. *Phytochem. Anal.* **6**:125-129.

140. Rawling, E.G., N.L. Martin, and R.E.W. Hancock. 1995. Epitope mapping of the *Pseudomonas aeruginosa* major outer membrane porin protein OprF. *Infect. Immun.* **63**:38-42.

141. Zhanell, G.G., J.A. Karlowsky, M.H. Saunders, R.J. Davidson, D.J. Hoban, R.E.W. Hancock, I. McLean, and L.E. Nicolle. 1995. Development of multiple-antibiotic-resistant (MAR) mutants of *Pseudomonas aeruginosa* after serial exposure to fluoroquinolones. *Antimicrob. Agents Chemother.* **39**:489-495.

142. Exner, M.M., P. Doig, T.J. Trust and R.E.W. Hancock. 1995. Isolation and characterization of a family of porin proteins from *Helicobacter pylori*. *Infect. Immun.* **63**:1567-1572.

143. Huang, H., D. Jeanteur, F. Pattus, and R.E.W. Hancock. 1995. Membrane topology and site-specific mutagenesis of *Pseudomonas aeruginosa* porin OprD. *Molec. Microbiol.* **16**:931-941.

144. Shang, E.S., M.M. Exner, T.A. Summers, C. Martinich, C.I. Champion, R.E.W. Hancock, and D.A. Haake. 1995. The rare outer membrane protein, OmpL1, of pathogenic *Leptospira* species is a heat-modifiable porin. *Infect. Immun.* **63**:3174-3181.

145. Blanco, D.R., C.I. Champion, M.E. Exner, H. Erdjument-Bromage, R.E.W. Hancock, P. Tempst, J.N. Miller, and M.A. Lovett. 1995. Porin activity and sequence analysis of a 31-kilodalton *Treponema pallidum* subsp. *pallidum* rare outer membrane protein (Tromp1). *J. Bacteriol.* **177**:3556-3562.

146. McCutcheon, A.R., T.E. Roberts, E. Gibbons, S.M. Ellis, L.A. Babiuk, R.E.W. Hancock, and G.H.N. Towers. 1995. Antiviral screening of British Columbian medicinal plants. *J. Ethnopharmacol.* **49**:101-110.

147. Wong, R.S.Y., R.A. Wirtz, and R.E.W. Hancock. 1995. *Pseudomonas aeruginosa* outer membrane protein OprF as an expression vector for foreign epitopes: the effects of positioning and length on the antigenicity of the epitope. *Gene*, **158**:55-60.

148. Liao, X., and R.E.W. Hancock. 1995. Cloning and characterization of the *Pseudomonas aeruginosa* *pbpB* gene encoding penicillin-binding protein 3. *Antimicrob. Agents Chemother.* **39**:1871-1874.

149. Matsuura, H., G. Saxena, S.W. Farmer, R.E.W. Hancock and G.H.N. Towers. 1995. Antibacterial and antifungal compounds from *Empetrum nigrum*. *Planta Medica*, **61**: 580.

150. Doig, P., M.M. Exner, R.E.W. Hancock, and T.J. Trust. 1995. Isolation and characterization of a conserved porin protein from *Helicobacter pylori*. *J. Bacteriol.* **177**:5447-5452.

151. Forst, S., J. Waukau, G. Leisman, M. Exner and R.E.W. Hancock. 1995. Functional and regulatory analysis of the OmpF-like porin, OpnP, of the symbiotic bacterium *Xenorhabdus nematophilus*. *Molec. Microbiol.* **18**:779-789.

152. Sukhan, A. and R.E.W. Hancock. 1995. Insertion mutagenesis of the *Pseudomonas aeruginosa* phosphate-specific porin OprP. *J. Bacteriol.* 177:4914-4920.

153. Lutwyche, P., M.M. Exner, R.E.W. Hancock and T.J. Trust. 1995. A conserved *Aeromonas salmonicida* porin provides protective immunity to rainbow trout. *Infect. Immun.* 63:3137-3142.

154. Matsuura, H., G. Saxena, S.W. Farmer, R.E.W. Hancock and G.H.N. Towers. 1996. An antibacterial thiophene from *Balsamorhiza sagittata*. *Planta Medica.* 62:65-66.

155. Liao, X., I. Charlebois, C. Ouellet, M.-J. Morency, K. Dewar, J. Lightfoot, J. Foster, R. Siehnel, H. Schweizer, J. Lam, R.E.W. Hancock and R.C. Levesque. 1996. Physical mapping of 32 genetic markers on the *Pseudomonas aeruginosa* PAO1 chromosome. *Microbiology*, 142: 79-86.

156. Saxena, G., S.W. Farmer, R.E.W. Hancock and G.H.N. Towers. 1996. Chlorochimaphilin: A new antibiotic from *Moneses uniflora*. *J. Natural Products*, 59:62-65.

157. Hancock, R.E.W. and F. Bellido. 1996. Antibacterial *in vitro* activity of fourth generation cephalosporins. *J. Chemotherapy* 8, Suppl. no. 2: 31-36.

158. Kondejewski, L.H., S.W. Farmer, D.S. Wishart, R.E.W. Hancock and R.S. Hodges. 1996. Gramicidin S is active against both gram-positive and gram-negative bacteria. *Int. J. Pept. Prot. Res.* 47:460-466.

159. Huang, H. and R.E.W. Hancock. 1996. The role of specific surface loop regions in determining the function of the imipenem-specific pore protein OprD of *Pseudomonas aeruginosa*. *J. Bacteriol.*, 178:3085-3090.

160. Matsuura, H., G. Saxena, S.W. Farmer, R.E.W. Hancock, and G.H.N. Towers. 1996. Antibacterial and antifungal polyine compounds from *Glehnia littoralis* ssp *Leiocarpa*. *Planta Medica*, 62:256-259.

161. Hancock, R.E.W. and T.J. Falla. 1996. Antimicrobial peptides: Broad-spectrum antibiotics from nature. *Clin. Microbiol. Infect.*, 1:226-229.

162. Wong, R.S.Y. and R.E.W. Hancock. 1996. The effect of the length of a malarial epitope on its antigenicity and immunogenicity in an epitope presentation system using the *Pseudomonas aeruginosa* outer membrane protein OprF as the carrier. *FEMS Microbiol. Lett.* 140:209-214.

163. Rehm, B.H.A. and R.E.W. Hancock. 1996. Membrane topology of the outer membrane protein OprH from *Pseudomonas aeruginosa*: PCR-mediated site-directed insertion and deletion mutagenesis. *J. Bacteriol.*, 178:3346-3349.

164. Falla, T.J., D.N. Karunaratne, and R.E.W. Hancock. 1996. Mode of action of the antimicrobial peptide indolicidin. *J. Biol. Chem.*, 271:19298-19303.

165. Sukhan A. and R.E.W. Hancock. 1996. The role of specific lysine residues in the passage of anions through the *Pseudomonas aeruginosa* porin OprP. *J. Biol. Chem.* 271:21239-21242.

166. Pumbwe, L., M.J. Everett, R.E.W. Hancock, and L.J.V. Piddock. 1996. Role of *gyrA* mutation and loss of OprF in the multiple antibiotic resistance (MAR) phenotype of *Pseudomonas aeruginosa* G49. *FEMS Microbiol. Lett.* 143: 25-28.

167. Gough, M., R.E.W. Hancock and N.M. Kelly. 1996. Anti-endotoxic potential of cationic peptide antimicrobials. *Infect. Immun.* 64: 4922-4927.

168. Kondejewski, L.H., Farmer, S.W., Wishart, D.S., Kay, C.M., Hancock, R.E.W. and R.S. Hodges. 1996. Effect of ring size of gramicidin S analogs on structure, antibacterial and hemolytic activity. *J. Biol. Chem.*, 271: 25261-25268.

169. Blanco, D.R., C.I. Champion, M.M. Exner, E.S. Shang, J.T. Skare, R.E.W. Hancock, J.N. Miller and M.A. Lovett. 1996. Recombinant *Treponema pallidum* rare outer membrane protein 1 (Tromp1) expressed in *Escherichia coli* has porin activity and surface antigenic exposure. *J. Bacteriol.* 178: 6685-6692.

170. Mahasreshti, P.J., G.L. Murphy, J.H. Wyckoff, S. Farmer, R.E.W. Hancock and A.W. Confer. 1997. Purification and partial characterization of the OmpA family of proteins of *Pasteurella haemolytica*. *Infect. Immun.* 65: 211-218.

171. Hancock, R.E.W. 1997. The bacterial outer membrane as a drug barrier. *Trends in Microbiol.*, 5: 37-42.

172. McCutcheon, A.R., R.W. Stokes, L.M. Thorson, S.M. Ellis, R.E.W. Hancock, and G.H.N. Towers. 1997. Anti-mycobacterial screening of British Columbian Medicinal Plants. *Intl. Journ. Pharmacog.* 35:77-83.

173. Bina, J.E., F. Nano and R.E.W. Hancock. 1997. Utilization of alkaline phosphatase fusions to identify secreted proteins, including potential efflux proteins and virulence factors from *Helicobacter pylori*. *FEMS Microbiol. Lett.* 148: 63-68.

174. Hancock, R.E.W. 1997. Peptide antibiotics. *The Lancet*, 349: 418-422.

175. Liao, J. and R.E.W. Hancock. 1997. Susceptibility to β -lactam antibiotics of *Pseudomonas aeruginosa* overproducing penicillin-binding protein 3. *Antimicrob Agents Chemother.* 41: 1158-1161.

176. Falla, T.J. and R.E.W. Hancock. 1997. Improved activity of a synthetic indolicidin analog. *Antimicrob. Agents Chemother.* 41: 771-775.

177. Hancock, R.E.W. 1997. Antibacterial Peptides and the outer membranes of Gram negative bacilli. *J. Med. Microbiol.* 46: 1-3.

178. Liao, X. and R.E.W. Hancock. 1997. Identification of a penicillin-binding protein 3 homologue, PBPC, in *Pseudomonas aeruginosa*: Gene cloning and growth phase dependent expression. *J. Bacteriol.* 179: 1490-6.

179. Champion, C.I., D.R. Blanco, M.M. Exner, H. Erdjument-Bromage, R.E.W. Hancock, P. Tempst, J.N. Miller and M.A. Lovett. 1997. Sequence analysis and recombinant expression of a 28-kilodalton *Treponema pallidum* subsp. *pallidum* rare outer membrane protein (Tromp2). *J. Bacteriol.* 179: 1230-1238.

180. Wong, K.K.Y., K.Poole, N.Gotoh, and R.E.W.Hancock. 1997. Influence of OprM expression on multiple antibiotic resistance in *Pseudomonas aeruginosa*. *Antimicrob.Aagents Chemother.* 41: 2009-2012.

181. Fidai, S., S. W. Farmer, and R.E.W. Hancock. 1997. Interaction of cationic peptides with bacterial membranes. *Meth. Molec. Biol.*, 78: 187- 204.

182. Kobaisy, M., Z.Abramowski, L.Lerner, G.Saxena, R.E.W.Hancock, G.H.N.Towers, D.Doxsee, and R.W.Stokes. 1997. Antimycobacterial polyynes of Devil'sClub (*Oplopanax horridus*), a north american native medicinal plant. *J. Natural Prod.* 60: 1210-1213.

183. Luo, Y., J.R. Glisson, M.W. Jackwood, R.E.W. Hancock, M. Bains, I.N. Cheng, and C. Wang. 1997. Cloning and characterization of the major outer membrane protein gene (OmpH) of *Pasteurella multocida* X-73. *J. Bacteriol.* 179: 7856-7864.

184. Hancock, R.E.W., and R.Lehrer. 1998. Cationic peptides: a new source of antibiotics. *Trends in Biotechnol.* 16: 82-88. (includes cover photo)

185. Davey, M.L., R.E.W.Hancock, and L.M. Mutharia. 1998. Influence of culture conditions on expression of the 40-kilodalton porin protein of *Vibrio anguillarum* serotype 02. *Appl. Environ. Microbiol.* 64: 138-146.

186. Hancock, R.E.W. 1998. The therapeutic potential of cationic peptides. *Expert Opinion Invest. Dis.* 7: 167-174.

187. Hancock, R.E.W., R.Alm, J.Bina, and T.Trust. 1998. *Helicobacter pylori*: A surprisingly conserved bacterium. *Nature Biotechnology* 16: 216-217.

188. Houston,M.E., L.H.Kondejewski, D.N.Karunaratne, M.Gough, S.Fidai, R.S.Hodes, and R.E.W.Hancock. 1998. Influence of preformed α -helix and α -helix induction on the activity of cationic antimicrobial peptides. *J.Peptide Res.* 52:81-88.

189. Rawling, E.G., F.S.L. Brinkman, and R.E.W. Hancock. 1998. Role of the carboxy-terminal half of *Pseudomonas aeruginosa* major outer membrane protein OprF in cell shape, growth in low-osmolarity medium, and peptidoglycan association. *J. Bacteriol.* 180:3556-3562.

190. Zhang, L., T.Falla, M.Wu, S.Fidai, J.Burian, W.Kay, and R.E.W.Hancock. 1998. Determinants of

recombinant production of antimicrobial cationic peptides and creation of peptide variants in bacteria. *Biochem. Biophys. Res. Commun.* **247**:674-680.

191. Dykes, G.A., R.E.W.Hancock, and J.W.Hastings. 1998. Structural variations in nisin associated with different membrane mimicking and pH environments. *Biochem. Biophys. Res. Commun.* **247**:723-727.

192. Wu, M., and R.E.W.Hancock. 1999. Interaction of the cyclic antimicrobial cationic peptide bactenecin with the outer and cytoplasmic membrane. *J. Biol. Chem.* **274**:29-35.

193. Hancock, R.E.W., and D.S.Chapple. 1999. Peptide antibiotics. *Antimicrob. Agents Chemother.* **43**:1317-1323.

194. Gensberg, K., A.W.Smith, F.S.L.Brinkman, and R.E.W.Hancock. 1999. Identification of oprG, a gene encoding a major outer membrane protein of *Pseudomonas aeruginosa*. *J. Antimicrob. Chemother.* **43**:607-608.

195. Wu, M., E.Maier, R.Benz, and R.E.W.Hancock. 1999. Mechanism of interaction of different classes of cationic antimicrobial peptides with planar bilayers and with the cytoplasmic membrane of *Escherichia coli*. *Biochemistry* **38**:7235-7242.

196. Wu, M., and R.E.W.Hancock. 1999. Improved derivatives of bactenecin, a cyclic dodecameric antimicrobial cationic peptide. *Antimicrob. Agents Chemother.* **43**:1274-1276.

197. Friedrich, C., M.G. Scott, N.Karunaratne, H.Yan, and R.E.W.Hancock. 1999. Salt-resistant alpha-helical cationic antimicrobial peptides. *Antimicrob. Agents Chemother.* **43**:1542-1548.

198. Scott, M.G., H.Yan, and R.E.W.Hancock. 1999. Biological properties of structurally related α -helical cationic antimicrobial peptides. *Infect. Immun.* **67**:2005-2009.

199. Zhang, L., R.Benz, and R.E.W.Hancock. 1999. Influence of proline residues on the antibacterial and synergistic activities of α -helical peptides. *Biochemistry* **38**:8102-8111.

200. Ochs, M.M., M.P.McCusker, M.Bains and R.E.W.Hancock. 1999. Negative regulation of the *Pseudomonas aeruginosa* outer membrane porin OprD selective for imipenem and basic amino acids. *Antimicrob. Agents Chemother.* **43**:1085-1090.

201. Kondejewski, L.H., M.Jelokhani-Niaraki, S.W.Farmer, B. Lix, C.M.Kay, B.D.Sykes, R.E.W.Hancock, and R.S.Hodges. 1999. Dissociation of antimicrobial and hemolytic activities in cyclic peptide diastereomers by systematic alterations in amphipathicity. *J. Biol. Chem.* **274**:13181-13192.

202. Brinkman, F.S.L., G. Schoofs, R.E.W.Hancock, and R.DeMot. 1999. Influence of a putative ECF sigma factor on expression of the major outer membrane protein, OprF, in *Pseudomonas aeruginosa* and *Pseudomonas fluorescens*. *J.Bacteriol.* **181**:4746-4754.

203. Ochs, M.M., C.-D. Lu, R.E.W.Hancock and A.T. Abdelal. 1999. Amino acid-mediated induction of the basic amino acid-specific outer membrane porin OprD from *Pseudomonas aeruginosa*. *J.Bacteriol.* **181**:5426-5432.

204. Macfarlane, E.L.A., A.Kwasnicka, M.M.Ochs, and R.E.W.Hancock. 1999. PhoP-PhoQ homologues in *Pseudomonas aeruginosa* regulate expression of the outer-membrane protein OprH and Polymyxin B resistance. *Molec. Microbiol.* **34**:305-316.

205. Scott, M.G., M.R.Gold, and R.E.W.Hancock. 1999. Interaction of cationic peptides with lipoteichoic acid and Gram positive bacteria. *Infect. Immun.* **67**:6445-6453.

206. Bina, J., R. A. Alm, M. Uria-Nuickelsen, S.R.Thomas, T.J.Trust, and R.E.W. Hancock. 2000. *Helicobacter pylori* uptake and efflux: Basis for intrinsic susceptibility to antibiotics in vitro. *Antimicrob. Agents Chemother.* **44**:248-254.

207. Scott, M.G.. A.C.E. Vreugdenhil, W.A. Buurman, R.E.W. Hancock, and M.R. Gold. 2000. Cutting Edge: Cationic Antimicrobial Peptides Block the Binding of Lipopolysaccharide (LPS) to LPS Binding Protein. *J. Immunol.* **164**: 549-553.

208. Bina, J., M.Bains, and R.E.W. Hancock. 2000. Functional expression in *Escherichia coli* and membrane topology of porin HopE, a member of a large family of conserved proteins in

Helicobacter pylori. J.Bacteriol. 182:2370-2375.

209. Jia, X., A. Patrzykat, R. Devlin, P.A. Ackerman, G.K. Iwama, and R.E.W. Hancock. 2000. Antimicrobial peptides protect Coho salmon from *Vibrio anguillarum* infections. Appl. Environ. Microbiol. 66:1928-1932.

210. Wong, K.K.Y., and R.E.W. Hancock. 2000. Insertion mutagenesis and membrane topology model of the *Pseudomonas aeruginosa* outer membrane protein OprM. J.Bacteriol. 182:2402-2410.

211. Chung, W., and R.E.W. Hancock. 2000. Action of lysozyme and nisin mixtures against lactic acid bacteria. Intl. J. Food Microbiol. 60:25-32.

212. Hancock, R.E.W., and M.G. Scott. 2000. The role of antimicrobial peptides in animal defences. Proc. Natl. Acad. Sci. USA. 97:8856-8861.

213. Johnstone, S.A., K. Gelmon, L.D. Mayer, R.E.W. Hancock, and M.B. Bally. 2000. In vitro characterization of the anticancer activity of membrane-active cationic peptides. I. Peptide-mediated cytotoxicity and peptide-enhanced cytotoxic activity of doxycydrubicin against wild-type and P-glycoprotein over-expressing tumor cell lines. Anti-Cancer Drug Design. 15:151-160.

214. Rosenberger, C.M., M.G. Scott, M.R. Gold, R.E.W. Hancock, and B.B. Finlay. 2000. *Salmonella typhimurium* and lipopolysaccharide induce similar changes in macrophage gene expression. J. Immunol. 164:5894-5904.

215. Friedrich, C.L., D. Moyles, T.J. Beveridge, and R.E.W. Hancock. 2000. Antibacterial action of structurally diverse cationic peptides on Gram-positive bacteria. Antimicrob. Agents Chemother. 44:2086-2092.

216. Ochs, M.M., M.Bains, and R.E.W. Hancock. 2000. Role of putative loops 2 and 3 in imipenem passage through the specific porin OprD of *Pseudomonas aeruginosa*. Antimicrob. Agents Chemother. 44:1983-1985.

217. Alm, R., J. Bina, B.M. Andrews, P. Doig, R.E.W. Hancock and T.J. Trust. 2000. Comparative genomics of *Helicobacter pylori*: Analysis of the outer membrane protein families. Infect. Immun. 68:4155-4168.

218. Hancock, R.E.W. 2000. Cationic antimicrobial peptides: towards clinical applications. Expert Opinion Invest. Dis. 9:1723-1729.

219. MacFarlane, E.L.A., A. Kwasnicka, and R.E.W. Hancock. 2000. Role of *Pseudomonas aeruginosa* PhoP-PhoQ in resistance to antimicrobial peptides and aminoglycosides. Microbiol. 146: 2543-2554.

220. Hancock, R.E.W., and D.P. Speert. 2000. Antibiotic resistance in *Pseudomonas aeruginosa*. Mechanisms and impact on treatment. Drug Resist. Updates. 3:247-255.

221. Stover, K.C., X.Q. Pham, A.L. Erwin, S.D. Mizoguchi, P. Warrener, M.J. Hickey, F.S.L. Brinkman, W. O. Hufnagle, D.J. Kowalik, M. Lagrou, R.L. Garber, L. Goltry, E. Tolentino, S. Westbroek-Wadman, Y. Yuan, L.L. Brody, S.N. Coulter, K.R. Folger, A. Kas, R. Lim, K. Smith, D. Spencer, G.K.-S. Wong, Z. Wu, I. Paulsen, J. Reizer, M.H. Saier, R.E.W. Hancock, S. Lory, and M.V. Olson. 2000. Complete genome sequence of *Pseudomonas aeruginosa*: an opportunistic pathogen. Nature 406:959-964.

222. Brinkman, F.S.L., M.Bains and R.E.W. Hancock. 2000. The amino terminus of *Pseudomonas aeruginosa* outer membrane protein OprF forms channels in lipid bilayer membranes: correlation with a three-dimensional model. J. Bacteriol. 182:5251-5255 (includes cover photo).

223. Brinkman, F.S.L., R.E.W. Hancock and K.C. Stover. 2000. Sequencing solutions: Use of volunteer annotators organized via the Internet. Nature 406:933.

224. Hancock, R.E.W. and G. Diamond. 2000. The role of cationic antimicrobial peptides in innate host defences. Trends Microbiol. 8:402-410.

225. Osusky, M., G. Zhou, L.Osuska, R.E.W. Hancock, W.W. Kay and S. Misra. 2000. Transgenic plants expressing cationic peptide chimeras exhibit broad-spectrum resistance to phytopathogens. Nature Biotech. 18:1162-1166.

226. Scott, M.G., C.M. Rosenberger, M.R. Gold, B.B. Finlay, and R.E.W. Hancock. 2000. An α -helical cationic antimicrobial peptide selectively modulates macrophage response to LPS and directly alters macrophage gene expression. *J. Immunol.* 165:3358-3365.

227. Jelokhani-Niaraki, M., L.H. Kondajewski, S.W. Farmer, R.E.W. Hancock, C.M. Kay, and R.S. Hodges. 2000. Diastereoisomeric analogues of gramicidin S: structure, biological activity and interaction with bilayers. *Biochem. J.* 349:747-755.

228. Scott, M.G., and R.E.W. Hancock. 2000. Cationic antimicrobial peptides and their multifunctional role in the immune system. *Crit. Rev. Immunol.* 20:407-431.

229. Zhang, L., P. Dhillon, H. Yan, S. Farmer, and R.E.W. Hancock. 2000. Interactions of bacterial cationic peptide antibiotics with outer and cytoplasmic membranes of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 44:3317-3321.

230. Zhang, L., M.G. Scott, H. Yan, L.D. Mayer, and R.E.W. Hancock. 2000. Interaction of polyphemusin I and structural analogs with bacterial membranes, lipopolysaccharide and lipid monolayers. *Biochem.* 39:14504-14514.

231. Rozek, A., C.L. Friedrich, and R.E.W. Hancock. 2000. Structure of the bovine antimicrobial peptide indolicidin bound to dodecylphosphocholine and sodium dodecyl sulfate micelles. *Biochem.* 39:15765-15774.

232. Prosser, R.S., P.A. Luchette, P.W. Westermann, A. Rozek, and R.E.W. Hancock. 2000. Determination of membrane immersion depth with O₂: A high pressure 19F NMR study. *Biophys. J.* 80:1406-1430.

233. Wong, K.K.Y., F.S.L. Brinkman, R.S. Benz, and R.E.W. Hancock. 2001. Evaluation of a structural model of *Pseudomonas aeruginosa* outer membrane protein OprM, an efflux component involved in intrinsic antibiotic resistance. *J. Bacteriol.* 183:367-374.

234. Brinkman, F.S.L., I. Wan, R.E.W. Hancock, A.M. Rose, and S.J. Jones. 2001. PhyloBLAST: A tool facilitating phylogenetic analysis of BLAST results. *Bioinformatics* 17:385-387.

235. Patrzykat, A., L. Zhang, V. Mendoza, G.K. Iwama, and R.E.W. Hancock. 2001. Synergy of histone-derived peptides of Coho salmon with lysozyme and the flounder peptide pleurocidin. *Antimicrob. Agents Chemother.* 45:1337-1342.

236. Yan, H., and R.E.W. Hancock. 2001. Synergistic interactions between mammalian antimicrobial defense peptides. *Antimicrob. Agents Chemother.* 45:1558-1560.

237. Friedrich, C.L., A. Rozek, A. Patrzykat, and R.E.W. Hancock. 2001. Structure and mechanism of action of an indolicidin peptide derivative with improved activity against Gram-positive bacteria. *J. Biol. Chem.* 276: 24015-24022.

238. Lange, C.F., R.E.W. Hancock, J. Samuel, and W.H. Finlay. 2001. In vitro aerosol delivery and regional airway surface liquid concentration of a liposomal cationic peptide. *J. Pharmaceut. Sci.* 90:1647-1657.

239. Hancock, R.E.W. 2001. Cationic peptides: effectors in innate immunity and novel antimicrobials. *Lancet Infectious Diseases* 1:156-164.

240. Brinkman, F.S.L., E.L.A. MacFarlane, P. Warrener, and R.E.W. Hancock. 2001. Evolutionary relationships amongst virulence associated histidine kinases. *Infect. Immun.* 69:5207-5211.

241. Zhang, L., A. Rozek, and R.E.W. Hancock. 2001. Interaction of cationic antimicrobial peptides with model membranes. *J. Biol. Chem.* 276:35714-35722.

242. Luchette, P.A., T.N. Vetman, R.S. Prosser, R.E.W. Hancock, M.P. Nieh, C.J. Glinka, S. Krueger, and J. Katsaras. 2001. Morphology of fast-tumbling bicelles: a small angle neutron scattering and NMR study. *Biochim. Biophys. Acta* 1513:83-94.

243. Chiu, C.H., S. Wong, R.E.W. Hancock, and D.P. Speert. 2001. Adherence of *Burkholderia cepacia* to respiratory tract epithelial cells and inhibition with dextrans. *Microbiol.* 147:2651-2658.

244. Hancock, R.E.W., and W.R. Strohl. 2001. Antimicrobials in the 21st Century. *Curr. Opin. Microbiol.* 4:491-492.

245. Hancock, R.E.W. 2001. A brief on bacterial biofilms. *Nature Genetics* 29:360. (News and views).

246. Desai, T.R., J.P. Wong, R.E.W. Hancock, and W.H. Finlay. 2002. A novel approach to the pulmonary delivery of liposomes in dry powder form to eliminate the deleterious effect of milling. *J. Pharmaceut. Sci.*, 91:482-91.

247. Hancock, R.E.W., and A. Rozek. 2002. Role of membranes in the activities of antimicrobial cationic peptides. *FEMS Microbiol. Lett.*, 206:143-149.

248. Hancock, R.E.W., and A. Patrzykat. 2002. Clinical development of cationic antimicrobial peptides: From natural to novel antibiotics. *Curr. Drug Targets – Infect. Disorders*. 2:79-83.

249. Patrzykat, A., C.L. Friedrich, L.Zhang, V. Mendoza, and R.E.W. Hancock. 2002. Sub-lethal concentrations of pleurocidin-derived antimicrobial peptides inhibit macromolecular synthesis in *Escherichia coli*. *Antimicrob. Agents Chemother.* 46:605-614.

250. Kondejewski, L.H., D.L. Lee, M. Jelokhani-Niaraki, S.W. Farmer, R.E.W. Hancock, and Robert S. Hodges. 2002. Optimization of microbial specificity in cyclic peptides by modulation of hydrophobicity within a defined structural framework. *J. Biol. Chem.* 277:67-74.

251. Hancock, R.E.W., and F.S.L. Brinkman. 2002. Function of *Pseudomonas* porins in uptake and efflux. *Annu. Rev. Microbiol.* 56:17-38.

252. Azghani, A., S. Idell, M. Bains, and R.E.W. Hancock. 2002. *Pseudomonas aeruginosa* outer membrane protein F is an adhesin in bacterial binding to lung epithelial cells in culture. *Microb. Pathogenesis* 33:109-114.

253. Gatto, N.T., S.M. Dabo, R.E.W. Hancock and A.W. Confer. 2002. Characterization of, and immune responses of mice to, the purified OmpA-equivalent outer membrane protein of *Pasteurella multocida* serotype A:3 (Omp28)., *Vet. Microbiol.* 87: 221-235.

254. Hirakata, Y., R. Srikumar, K. Poole, N. Gotoh, T. Suematsu, S. Kohno, S. Kamihira, R.E.W. Hancock, and D.P. Speert. 2002. Multidrug efflux systems play an important role in the invasiveness of *Pseudomonas aeruginosa*. *J. Exp. Med.* 196:109-118.

255. Roy, S., H.G. Lombart, W.D. Lubell, R.E.W. Hancock, and S.W. Farmer. 2002. Exploring relationships between mimic configuration, peptide conformation and biological activity in indolizidin-2-one amino acid analogs of gramicidin S. *J. Pept. Res.* 60:197-213.

256. Brinkman, F.S.L., J.L. Blanchard, A. Cherkasov, H. Greberg, Y. Av-Gay, R.C. Brunham, R.C. Fernandez, B.B. Finlay, S.P. Otto, B.F.F. Ouellette, P.J. Keeling, A.M. Rose, R.E.W. Hancock and S.J.M. Jones (2002). Evidence that plant-like genes in *Chlamydia* species reflect an ancestral relationship between *Chlamydiaceae*, cyanobacteria and the chloroplast. *Genome Research*. 12:1159-1167.

257. Scott, M.G., D.J. Davidson, M.R. Gold, D. Bowdish, and R.E.W. Hancock. 2002. The human antimicrobial peptide, LL-37, is a multifunctional modulator of innate immune responses. *J. Immunol.* 169:3883-3891.

258. Yoon, S.S., R.F. Hennigan, G.M. Hilliard, U.A.Ochsner, K.Parvatiyar, M.C.Kamani, H.L.Allen, T.R.DeKievit, P.R. Gardner, U.Schwab, J.J.Rowe, B.H.Iglewski, T.R.McDermott, R.P.Mason, D.J.Wozniak, R.E.W. Hancock, M.R.Parsek, T.L.Noah, R.C.Boucher, and D.J.Hassett. 2002. *Pseudomonas aeruginosa* anaerobic respiration in biofilms: Relationships to cystic fibrosis pathogenesis. *Developmental Cell*. 3:593-603.

259. Lupp, C., R.E.W. Hancock, and E.G. Ruby. 2002. The *Vibrio fischeri sapABCDF* locus is required for normal growth, both in culture and in symbiosis. *Arch. Microbiol.* 179:59-65.

260. Desai, T.R., R.E.W. Hancock, and W.H. Finlay. 2002. A facile method of delivery of Liposomes by nebulization. *J. Controlled Release* 84:69-78.

261. Jo, J.T.H., F.S.L. Brinkman, and R.E.W. Hancock. 2003. Aminoglycoside efflux in *Pseudomonas aeruginosa*: involvement of novel outer membrane proteins. *Antimicrob. Agents Chemother.* 47:1101-1111.

262. Tamber, S., and R.E.W. Hancock. 2003. On the mechanism of solute transport in *Pseudomonas*.

Frontiers in Bioscience. 8:S472-S483.

263.Boehr, D.D., K. Draker, K. Koteva, M. Bains, R.E. Hancock, and G.D. Wright. 2003 Broad-spectrum peptide inhibitors of aminoglycoside resistance enzymes. *Chem. Biol.* 10:189-196.

264.Llamas, M.A., J.J. Rodriguez-Herva, R.E.W. Hancock, W. Bitter, J. Tommassen, and J.L. Ramos. 2003. Role of *Pseudomonas putida* tol-oprL gene products in uptake of solutes through the cytoplasmic membrane. *J. Bacteriol.* 185:4707-16.

265.McPhee, J.B., S. Lewenza and R.E.W. Hancock. 2003. Cationic antimicrobial peptides activate a two-component regulatory system, PmrA-PmrB, that regulates resistance to polymyxin B and cationic antimicrobial peptides in *Pseudomonas aeruginosa*. *Molec. Microbiol.* 50:205-219.

266.Haines, L.R., R.E.W. Hancock and T.W. Pearson. 2003. Cationic antimicrobial peptide killing of African trypanosomes and *Sodalis glossinidius*, a bacterial symbiont of the insect vector of sleeping sickness. *Vector Borne and Zoonotic Dis.* 3:175-186.

267.Hancock, R.E.W. 2003. Concerns regarding resistance to self-proteins (commentary). *Microbiol.* 149,3343-3344.

268.Rozek A., J.P. Powers, C.L. Friedrich, R.E.W. Hancock. 2003. Structure-based design of an indolicidin peptide analogue with increased protease stability. *Biochemistry.* 42:14130-14138.

269.Halevy, R., A. Rozek, S. Kolusheva, R.E.W. Hancock, and R. Jelinek. 2003. Membrane binding and permeation by indolicidin analogs studied by a biomimetic lipid/polydiacetylene vesicle assay. *Peptides* 24:1753-1761.

270.Powers, J.-P.S., and R.E.W. Hancock. 2003. The relationship between peptide structure and antibacterial activity. *Peptides* 24:1681-1691.

271.Anderson, R.C., R.E.W. Hancock and P.-L. Yu. 2003. Antimicrobial activity and bacterial membrane interaction of ovine-derived cathelicidins. *Antimicrob. Agents Chemother.* 48:673-676.

272.Desai, T.R., R.E.W. Hancock, and W.H. Finlay. 2003. Delivery of liposomes in dry powder form: aerodynamic dispersion properties. *Europ. J. Pharmaceut. Sci.* 20:459-467.

273.Davidson, D.J., A.J. Currie, G. Reid, D.M.E. Bowdish, K. MacDonald, R. Ma, R.E.W. Hancock, and D.P. Speert. 2004. The cationic antimicrobial peptide LL-37 modulates dendritic cell development and DC-induced T cell polarisation. *Journal of Immunology.* 172:1146-1156.

274.Powers, J.P., A. Rozek, and R.E.W. Hancock. 2004. Structure-activity relationships for the β -hairpin cationic antimicrobial peptide polyphemusin I. *Biochim. Biophys. Acta*, in press.

275.Hokamp K., F.M. Roche, M. Acab, M.E. Rousseau, B. Kuo, D. Goode, D. Aeschliman, J. Bryan, L. Babiuk, R.E.W. Hancock, and F.S.L. Brinkman. 2004. ArrayPipe: a flexible processing pipeline for microarray data. *Nucl. Acids Res.*, in press.

276.Bowdish D.M.E., D.J. Davidson, D.P. Speert, and R.E.W. Hancock. 2004. The human cationic peptide LL-37 induces activation of the extracellular signal regulated kinase and p38 kinase pathways in primary human monocytes. *J. Immunol.* 172:3758-3765.

277.Lee, D.L.,J.P.S. Powers, K. Pflegerl, M.L. Vasil, R.E.W. Hancock, and R.S. Hodges. 2004. Effects of single D-amino acid substitutions on disruption of β -sheet structure and hydrophobicity in cyclic 14-residue antimicrobial peptide analogs related to gramicidin S. *J. Peptide Res.* 63:69-84.

278.Finlay, B.B., and R.E.W. Hancock. 2004. Can innate immunity be enhanced to treat infections? *Nature Microbiol. Rev.*, in press.

279.Roche, F.M. K. Hokamp, M. Acab, L.A. Babiuk, R.E.W. Hancock, and F.S.L. Brinkman. 2004. ProbeLynx: A tool for updating the association of microarray probes to genes. *Nucl. Acids Res.* In press.

280.Bowdish, D.M.E., D.J. Davidson, and R.E.W. Hancock. 2004. The role of host defence and related peptides in immunity. *Current Protein & Peptide Science*, in press.

281.McPhee, J.B. M.G. Scott, and R.E.W. Hancock. 2004. Design of host defence peptides for antimicrobial and immunity enhancing activities. *Combinat. Chem. High Throughput Screen.* in press.

282. Jung, D., A. Rozek, M. Okon, and R.E.W. Hancock. 2004. Structural transitions as determinants of the calcium dependent antibiotic daptomycin. *Chem. Biol.*, In press.

283. Osusky, M. L. Osuska, R. E. Hancock, W. W. Kay, and S. Misra. 2004. Transgenic potatoes expressing a novel cationic peptide are resistant to late blight and pink rot. *Transgenic Research* PC1223:1-10, in press.

BOOK CHAPTERS AND REVIEWS

1. Hancock, R.E.W., R.A. Skurray and P. Reeves. 1975. A mutation in *Escherichia coli* K-12 affecting conjugation, phage resistance and a major membrane protein. *Proc. Aust. Biochem. Soc.* 7:80.
2. Braun, V., R.E.W. Hancock, K. Hantke and A. Hartman. 1976. Functional organization of the outer membrane of *Escherichia coli*: Phage and colicin receptors as components of iron uptake systems. In *Progress in Clinical and Biological Research* 17:11-32 (Revel, J.P., U. Henning and C.F. Fox, eds.) Alan R. Liss Inc., New York.
3. Hancock, R.E.W. 1981. Aminoglycoside uptake and mode of action - with special reference to streptomycin and gentamicin. I. Antagonists and mutants. *J. Antimicrob. Chemother.* 8:249-276.
4. Hancock, R.E.W. 1981. Aminoglycoside uptake and mode of action - with special reference to streptomycin and gentamicin. II. Effects of aminoglycosides on cells. *J. Antimicrob. Chemother.* 8:429-445.
5. Benz, R., R.E.W. Hancock and T. Nakae. 1982. Porins from gram-negative bacteria in lipid bilayer membranes. In "Transport in Biomembranes: Model systems and reconstitution" (Antolini et al, eds), pp. 123-134. Raven Press, New York.
6. Hancock, R.E.W., B. Loh and T.I. Nicas. 1983. Molecular Interactions of aminoglycosides with bacteria - Implications for therapy. *Proc. 13th International Congress of Chemotherapy* 35:1-3.
7. Hancock, R.E.W. and T.I. Nicas. 1984. Resistance to antibacterial agents acting on cell membranes. In "Antimicrobial Drug Resistance" (Bryan, L.E., ed). pp.147 - 171. Academic Press, New York.
8. Benz, R., K. Poole and R.E.W. Hancock. 1984. In Ionic channels in membranes. *Biophysical Discussions*. (Parsegian, V.A., ed.). pp. 81-82. Rockefeller University Press, New York.
9. Hancock, R.E.W. 1984. Alterations in outer membrane permeability. *Annu. Rev. Microbiol.* 38:237-264.
10. Mutharia, L.M., J.S.L. Lam and R.E.W. Hancock, 1985. Use of monoclonal antibodies in the study of common antigens in gram negative bacteria. In (A.J.L. Macario and E.C. de Macario, eds.) "Monoclonal antibodies against bacteria", pp. 131-142, Academic Press, New York.
11. Lam, J.S.L., L.M. Mutharia and R.E.W. Hancock. 1985. Application of monoclonal antibodies to the study of the surface antigens in *Pseudomonas aeruginosa*. In (A.J.L. Macario and E.C. Macario, eds.) "Monoclonal antibodies against bacteria", Vol. II, pp. 143-157, Academic Press, New York.
12. Hancock, R.E.W. 1985. The role of the cell surface components of *Pseudomonas aeruginosa* in virulence. In (G.G. Jackson and J. Thomas, eds.) *Bayer Symposium VII: Pathogenesis of Bacterial Infections*. pp. 247-256. Springer-Verlag, Berlin.
13. Hancock, R.E.W. 1985. Monoclonal antibody protection against *Pseudomonas aeruginosa*. In (G.G. Jackson and J. Thomas, eds.) pp. 422-424. *Bayer Symposium VII: The Pathogenesis of Bacterial Infections*. Springer-Verlag, Berlin.
14. Hancock, R.E.W. 1985. Laboratory characterization of lipopolysaccharides (LPS). In (G.G. Jackson and J. Thomas, eds.) p. 398. *Bayer Symposium VII: The Pathogenesis of Bacterial Infections*. Springer-Verlag, Berlin.
15. Hancock, R.E.W. 1985. The *Pseudomonas aeruginosa* outer membrane permeability barrier and how to overcome it. In (Speert, D.P., and R.E.W. Hancock, Ed.) *Antibiotics and Chemotherapy*, pp. 95-102, S. Karger, Basel.37.
16. Hancock, R.E.W. and L.M. Mutharia. 1985. Monoclonal Antibodies against bacterial outer membrane antigens. In *Advances in Experimental Medicine and Biology*, Vol. 185 (Atassi, M. and H.L. Bachrach, eds.) pp. 215-222. Plenum Press Inc., New York.
17. Nikaido, H. and R.E.W. Hancock. 1986. Outer membrane permeability of *Pseudomonas aeruginosa*. In (Sokatch, J.R., ed.) *The Bacteria: A treatise on structure and function*. X. pp. 145-193. Academic Press, London.

18. Hancock, R.E.W., E.C.A. Mouat, S.L. Butters and D.P. Speert. 1986. The outer membrane proteins of *Pseudomonas aeruginosa*: Immunotherapeutic potential. In (Robbins, J.B., ed.) *Seminars in Infectious Diseases*.
19. Moore, R.A. and R.E.W. Hancock. 1985. The penetration of antibiotics into the antibiotic resistant bacterium *Pseudomonas aeruginosa*: Interaction of polycationic antibiotics at a common site on the lipopolysaccharide. *Chemioterapia*. 4, Suppl. No. 2: 3-5.
20. Hancock, R.E.W. 1987. Model membrane studies of porin function. In *Bacterial Outer Membranes as Model Systems* (Inouye, M., ed.) pp. 187-225. John Wiley and Sons, New York.
21. Hancock, R.E.W., E.A. Worobec, K. Poole and R. Benz. 1987. Phosphate-binding site of *Pseudomonas aeruginosa* outer membrane protein P. In (Torriani, A., Silver, S., Rathman, F., Wright, A., and Yagel, E., eds.) "Phosphate metabolism and cellular regulation in microorganisms." pp.176-180, ASM, Washington.
22. Moore, R.A., W.A. Woodruff and R.E.W. Hancock. 1987. Antibiotic uptake pathways across the outer membrane of *Pseudomonas aeruginosa*. *Antibiotics and Chemotherapy*, 39:172-181.
23. Hancock, R.E.W. and N. Kelly. 1987. Role of outer membrane components in *Pseudomonas aeruginosa* colonization of the cystic fibrosis lung. *Pediatr. Pulmon.*, Suppl. 1:89-91.
24. Hancock, R.E.W. and W.A. Woodruff. 1988. Roles of porin and β -lactamase in β -lactam resistance of *Pseudomonas aeruginosa*. *Rev. Infect. Dis.* 10:770-775.
25. Hancock, R.E.W. and A. Bell. 1989. Antibiotic uptake into Gram negative bacteria. In "Perspectives in anti-infective therapy" (G.G. Jackson, H.D. Schlumberger, and H.J. Zeiler, eds.) pp. 21-28. Vieweg and Sohn, Braunschweig.
26. Martin, N.L. and R.E.W. Hancock. 1990. Function and structure of the major components of the outer membrane of gram-negative bacteria. In *Advances in Brucellosis Research* (ed. L.G. Adams), pp. 55-75, Texas A & M University Press, College Station.
27. Siehnel, R.J., N.L. Martin and R.E.W. Hancock. 1990. Function and structure of the porin proteins OprF and OprP of *Pseudomonas aeruginosa*. In *Pseudomonas: Biotransformations pathogenesis and evolving biotechnology*. Silver, S., A.M. Chakrabarty, B. Iglesias, and S. Kaplan. pp. 328-342. American Society for Microbiology, Washington.
28. Hancock, R.E.W. and N. Karunaratne. 1990. LPS integration into outer membrane structures. In *Cellular and Molecular Aspects of Endotoxin Reactions*. Vol. 1, pp. 191-196. (Nowotny, A., Spitzer, J.J., and Ziegler, E.J., eds.) Elsevier Science Publishers B.V., Amsterdam.
29. Hancock, R.E.W. 1991. Bacterial outer membranes: Evolving Concepts. *ASM News* 57:175-182.
30. Bellido, F., R.L. Finnen, N.L. Martin, R.J. Siehnel and R.E.W. Hancock. 1992. Function and structure of *Pseudomonas aeruginosa* outer membrane protein OprF. In: *Pseudomonas: Molecular Biology and Biotechnology*. (Galli, E., S. Silver and B. Witholt, eds.). pp. 170-176. American Society for Microbiology, Washington, D.C.
31. Hancock, R.E.W. and M.H. Brown. 1992. Bacterial Porins. *Today's Life Science* 4:24-32.
32. Bellido, F., and R.E.W. Hancock. 1994. Susceptibility and resistance of *Pseudomonas aeruginosa* to antimicrobial agents. In *Pseudomonas aeruginosa* as an opportunistic pathogen. (Campa, M., M. Bendinelli, and H. Friedman, eds) pp.321-348. Plenum Press, NY.
33. Hancock, R.E.W., C. Egli and N. Karunaratne. 1994. Molecular organization and structural role of outer membrane macromolecules. In *Bacterial Cell Envelope* (J.M. Ghysen and Hakenbeck, R., eds). pp. 263-279. pp. 263-279. Elsevier Science Publishers BV, Amsterdam.
34. Hancock, R.E.W., and K. Piers. 1994. Outer membrane proteins. In *Escherichia coli* in animals. (Gyles C.L., ed.). pp. 495-532. CAB International, Wallingford, U.K.
35. Siehnel, R.J., R.S.Y. Wong, H. Huang and R.E.W. Hancock 1994. Cloning of outer membrane protein genes and molecular genetic methods for studying structure function relationships. *Meth. Molec. Genet.* 3:311-324.
36. Hancock, R.E.W. and T.E. McCurry. 1993. Canadian Bacterial Diseases Network: a new approach to University-Industry relationships. *Clin. Invest. Med.* 16:306-313.
37. Hancock, R.E.W., T. Falla and M.H. Brown, 1995. Cationic Bactericidal peptides. *Adv. Microb. Physiol.* 37:135-175.
38. Hancock, R.E.W. 1994. Bacterial transport as an import mechanism and target for antimicrobials. In Georgopapadakou, N. ed., "Drug transport in antimicrobial and anticancer chemotherapy", pp. 289-306. Marcel Dekker, Inc., New York.
39. Hancock, R.E.W., S. Fidai, M. Gough, M. Wu, and K. Piers. 1994. Small Peptide Antibiotics.

Pediatric Pulmonology 10(S):161-162.

40. Hancock, R.E.W., and T. Falla, 1996. Cationic Peptides. In Biotechnology of Antibiotics, Strohl, W.R., 2nd Edition, Ch. 16: 467-492. Marcell Dekker Inc., N.Y.
41. Hancock, R.E.W., K. Piers, M. Brown, T. Falla, M. Gough, M. Wu and S. Fidai. 1996. Cationic peptides: a class of antibiotics able to access the self promoted uptake pathway across the outer membrane of *Pseudomonas aeruginosa*. In: Molecular Biology of Pseudomonads. (Nakazawa, T., Furukawa, K., Haas, D., and Silver, S., eds) ASM Press, Washington.
42. Hancock, R.E.W., and D.P. Speert. 1996. Antibiotics for *Pseudomonas* and related infections. In Cystic Fibrosis - current topics (Dodge, J.A., Brock, D.J.H., and Widdicombe, J.H., eds.) Vol. 3, pp. 245-266. John Wiley and Sons Ltd.
43. Hancock, R.E.W. and M. Exner. 1997. Isolation and characterization of porins from *H. pylori*. In Clayton, C.L. and H.L.T. Mobley eds. Methods Molec. Med. *Helicobacter pylori* protocols. pp. 191-204. Humana Press In., Totowa, N.J.
44. Hancock, R.E.W. and R. Wong. 1997. Potential of protein OprF of *Pseudomonas* in bivalent vaccines. In Domdey, H., W. Lubitz, F. Schodel and B. von Specht eds. Behring Institute Mitteilungen Vol. 98, 283-290. DMV, Marburg.
45. Hancock, R.E.W. 1997. The role of fundamental research and biotechnology in finding solutions to the global problem of antibiotic resistance. Clin. Infect. Dis., 24 (Supplement 1): S148-S150.
46. Hancock, R.E.W., and E.A. Worobec. 1998. Outer membrane proteins, In (Montie T., ed.) Biotechnology Handbooks. 10 *Pseudomonas*, pp. 139-167. Plenum Press, London.
47. Hancock, R.E.W. 1998. Resistance mechanisms in *Pseudomonas aeruginosa* and other nonfermentative Gram-negative bacteria. Clin. Infect. Dis. 27: S93-99.
48. Hancock, R.E.W. and J.S. Lam. 1998. *Pseudomonas aeruginosa*: Infection and Immunity. In (P.J.Delves ed.) Encyclopaedia of Immunology, 2nd Edition. 4:2042-2045. Academic Press, London.
49. Hancock, R.E.W., and D.Knowles. 1998. Are we approaching the end of the antibiotic era? Editorial overview. Curr. Opinion Microbiol. 1:493-494.
50. Towers, G.H.N., McCutcheon, A., Saxena, G., Matsuura, H., Page, J., Farmers, S., Gibbons, E. Roberts, T.E., Babiuk, L.A., Thorson, I.M., Stokes, R.W., Sokol, P., Klingemann, H., and Hancock, R.E.W. 1999. Anti-microbial activities of phytochemicals from British Columbian medicinal plants. In: Chemistry, Biological, and Pharmacological Properties of Medicinal Plants from the Americas. Ed. Hostettmann, K., Gupta, M.P., and Marston, A. Harwood Academic Publ. Switzerland. pp. 125-142.
51. Hancock, R.E.W. 1999. Host defence (cationic) peptides: What is their future clinical potential? Drugs 57: 469-473.
52. Hancock, R.E.W., and A.Patrzykat. 1999. Cationic peptide antibiotics: a diversified portfolio for the future. Chimica Oggi (Biocides Today supplement) pp. 20-21.
53. Hancock, R.E.W., M. Scott, C.Friedrich, L.Zhang, H.Yan, P.Nair, A.Patrzykat, and A.Rozek. 1999. Cationic peptide antibiotics for use in treatment of *Pseudomonas aeruginosa* infections of cystic fibrosis patients. Clin. Microbiol. Infect. 5:S52-S53.
54. Zhang, L., and R.E.W.Hancock. 2000. Peptide antibiotics. In Hughes, D., and Andersson, D.I., eds. Antibiotic resistance and antibiotic development. pp. 209-232. Harwood Academic Publishers.
55. Hetru, C., J.A.Hoffmann, and R.E.W.Hancock. 2000. Insect cationic antimicrobial peptides. In Dutton, C., M.Haxell, H.I. McArthur, and R. Wax. Peptide antibiotics: Discovery, modes of action and applications. Marcel Dekker, New York.
56. Macfarlane, E.L.A., and R.E.W. Hancock. 2000. Antibiotic resistance and survival in the host. In (Brogden, K., J.A. Roth, T.B. Stanton, C.A. Bolin, F.C. Minion, and M.J. Wannemuehler, eds.) Virulence mechanisms of bacterial pathogens, 3rd edition, pp. 93-104, ASM Press, Washington, DC.
57. Hancock, R.E.W., and P.Nair. 2000. Cationic antimicrobial peptide antibiotics. Curr. Opin. Antiinfective Invest. Drugs. 2:140-148.
58. Hancock, R.E.W. 2001. Mechanism of action of imipenem and relevance of porin OprD. Infections in Oncology 5:2-3.
59. Hancock, R.E.W., and A. Patrzykat. 2001. Antimicrobial peptides for fish disease control. In "Recent advances in marine biotechnology" Vol. 7: Seafood Safety and Human Health, Oxford and IBH Publishing Co., New Delhi.

60. Lewenza, S., and R.E.W. Hancock. 2001. Post-genomic *Pseudomonas*. *Genome Biology* 3:4002.1-4002.2.
61. Hancock, R.E.W. 2002. Cationic anti-microbial peptides. *Biochem. Cell Biol.* 80:140.
62. Devine, D.A., and R.E.W. Hancock. 2002. Cationic peptides: Distribution and mechanisms of resistance. *Curr. Pharmaceut. Design* 8:99-110.
63. Tamber, S., and R.E.W. Hancock. 2003. Electrophoresis and blotting of DNA. In *Nature Encyclopaedia of Life Sciences*. London: <http://www.els.net/> [<http://dx.doi.org/10.1038/npg.els.0003746>].
64. Hancock, R.E.W., and S. Tamber. 2004. Porins of the Outer Membrane of *Pseudomonas aeruginosa*. In R. Benz, ed., "Structure and Function of Bacterial and Eukaryotic Porins". John Wiley & Sons, NJ.
65. Hassett, D.J., P.A. Limbach, R.F. Hennigan, K.E. Klose, R.E.W. Hancock, M.D. Platt, D.F. Hunt. 2003. Bacterial biofilms of importance to medicine and bioterrorism: proteomic techniques to identify novel vaccine components and drug target. *Exp. Opin. Biol. Ther.* 3: 1201-1207.
66. Tamber, S., and R.E.W. Hancock. 2004. The Outer Membranes of Pseudomonads. In J.L. Ramos, ed., *The Pseudomonads*, Vol. I. Pp. 575-601. Kluwer Academic, NY.
67. Hancock, R.E.W., and D.A. Devine, editors. 2004. Mammalian host defence peptides. *Advances in Molecular and Cellular Microbiology Series*. Cambridge University Press, NY.
68. Hancock, R.E.W., and D.A. Devine. 2004. "Antimicrobial" or "host defence" peptides. In, D.A. Devine and Hancock, R.E.W., eds. *Mammalian host defence peptides*. pp. 1-4 Cambridge University Press, NY.
69. Hancock, R.E.W. 2004. Bacterial Structure and Physiology: Influence on Susceptibility to Cationic Antimicrobial Peptides. In D.A. Devine and Hancock, R.E.W., eds. *Mammalian host defence peptides*. pp. 229-244. Cambridge University Press, NY.
70. Hancock, R.E.W., and S. Tamber. 2004. Porins of the outer membrane of *Pseudomonas aeruginosa*. In Benz, R., ed. *Bacterial and Eukaryotic Porins*. Ch. 4, pp. 61-77, Wiley VCH Verlag, Weinheim.

ISSUED PATENTS

1. Cationic peptides and method of production. R.E.W. Hancock, M.H. Brown, and K.L. Piers. **US Patent No 5,593,866, issued Jan 14, 1997.**
2. CEMA cationic peptide and polynucleotides encoding CEMA. R.E.W. Hancock, M.H. Brown, and K.L. Piers. **US patent No. 5,707,855, issued Jan 13, 1998.**
3. Treatment of endotoxin associated disorders with cationic peptides. R.E.W. Hancock, M.H. Brown, K.L. Piers, and N.Kelly. **US patent No 5,688,767, issued Nov 18, 1997.**
4. Treatment of endotoxin associated disorders with cationic peptides. R.E.W. Hancock, M.H. Brown, K.L. Piers, and N.Kelly. **US patent No 5,789,377, issued August 4, 1998.**
5. Antimicrobial cationic peptides. R.E.W. Hancock and N.Karunaratne. **US Patent No 5,877,274, issued March 2, 1999.**
6. Antimicrobial cationic peptides. R.E.W. Hancock and N.Karunaratne. **US Patent No 6,040,435 issued March 21, 2000.**
7. Antimicrobial cationic peptides. R.E.W. Hancock and N.Karunaratne. **US Patent No 6,057,291 issued May 2, 2000.**
8. Antimicrobial cationic peptides. R.E.W. Hancock and N.Karunaratne. **US Patent No 6,297,215 issued Oct 2, 2001.**
9. Antimicrobial cationic peptides. R.E.W. Hancock and N.Karunaratne. **US Patent No 6,465,429 issued Oct 15, 2002.**
10. Antimicrobial cationic peptides and methods of screening for the same. T. Falla, Hancock, R.E.W., and M. Gough. **US patent No 6,191,254, issued Feb 20, 2001.**
11. Antimicrobial peptide composition and method of screening for same. N.A.H. Lewis-Ruthven, R.S. Hodges, E.J. Prenner, D.S. Wishart, R.E.W. Hancock, L.H. Kondejewski and R.N. McElhaney **US patent No 6,358,921 issued March 19, 2002.**
12. Antimicrobial cationic peptide derivatives of Bactenecin. R.E.W. Hancock and M. Wu, **US Patent No 6,172,185 issued Jan 6, 2001.**
13. Anti-endotoxic, cationic antimicrobial peptides and methods of use therefore. R.E.W. Hancock, M. Gough, A. Patrzykat, X.Jia, and D.Woods, **US patent 6,288,212 issued September 11, 2001.**

14. Anti-endotoxic, cationic antimicrobial peptides and methods of use therefore. R.E.W.Hancock, M.Gough, A.Patrzykat, X.Jia, and D.Woods. **Australian Patent 758698. issued Sept. 27,2003.**
15. Antimicrobial peptides and methods of use thereof. R.E.W.Hancock and L.Zhang. **US patent 6,337,317 issued January 8, 2002.**
16. Antimicrobial peptides and methods of use thereof. R.E.W.Hancock and L.Zhang. **European patent 1,294,745 issued March 26, 2002.**



TRANSGENIC PLANTS THAT ARE RESISTANT TO A BROAD SPECTRUM OF PATHOGENS

ABSTRACT

5 Transgenic plants that express dermaseptin and/or temporin peptides are disclosed. In certain embodiments, these plants have enhanced, broad-spectrum pathogen resistance and are useful as agricultural or horticultural crops. In other embodiments, the plants are used to produce large quantities of the dermaseptin and/or temporin peptides.